NEUROPHOTONICS

REVIEW

Transcranial photobiomodulation for brain diseases: review of animal and human studies including mechanisms and emerging trends

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ABSTRACT. The brain diseases account for 30% of all known diseases. Pharmacological treatment is hampered by the blood–brain barrier, limiting drug delivery to the central nervous system (CNS). Transcranial photobiomodulation (tPBM) is a promising technology for treating brain diseases, due to its effectiveness, non-invasiveness, and affordability. tPBM has been widely used in pre-clinical experiments and clinical trials for treating brain diseases, such as stroke and Alzheimer's disease. This review provides a comprehensive overview of tPBM. We summarize emerging trends and new discoveries in tPBM based on over one hundred references published in the past 20 years. We discuss the advantages and disadvantages of tPBM and highlight successful experimental and clinical protocols for treating various brain diseases. A better understanding of tPBM mechanisms, the development of guidelines for clinical practice, and the study of dose-dependent and personal effects hold great promise for progress in treating brain diseases.

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1 Introduction

The brain is a vital component of the central nervous system (CNS) and is in charge of an individual's life activities, such as thinking, emotion, and memory. Brain regions are anatomically interconnected, exhibiting both specialization and collaboration in their functions, making it one of the most intricate and advanced systems found in nature. However, the brain is extremely vulnerable to various disorders, including cerebrovascular diseases, neurodegenerative diseases, brain inflammation, etc.,^{1–4} which have posed a significant burden on individuals, families, and society as a whole.

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Drug therapy is the most common therapeutic approach for brain diseases in clinical practice. However, drug therapy may produce adverse side effects, and the highly selective permeability of the blood–brain barrier (BBB) significantly limits brain drug delivery, resulting in a disappointing therapeutic effect.⁵ Some novel drug delivery strategies, such as viral vectors and non-viral nanoparticles, have demonstrated prodigious potential for delivering therapeutic to the brain. However, the safety of injecting a virus and the penetrability of the nanoparticles and their encapsulated payloads require further study.⁶ Recently, erythropoietin (EPO), stem cell therapy, transcranial magnetic stimulation (TMS), and transcranial direct current stimulation (tDCS) have been proposed as potential therapeutics for brain diseases.^{7–10} However, EPO naturally increases the hematocrit, which could heighten the risk of cardiovascular reperfusion.¹¹ For stem-based therapies, the ethical issues and safety concerns need further discussion.¹² TMS and tDCS lack clear mechanisms underlying their therapeutic effects, and the absence of widely accepted guidelines and standards for treating brain diseases impedes their clinical applications.^{13,14} Therefore, development of a safe, effective, and widely used therapeutic approach for brain diseases is urgent.

Transcranial photobiomodulation (tPBM), a non-invasive and non-thermal brain stimulation therapy, was proposed over 50 years ago. Numerous related devices are also available in the market. tPBM was known as low-level laser (light) therapy at the early stage, but more and more researchers prefer tPBM in recent years because it is more indicative of its scientific principle.¹⁵ tPBM refers to applying low irradiance (0.01 to 10 W/cm²) red to near-infrared (NIR) (600 to 1300 nm) light^{16,17} through the skull directly to brain tissue to achieve neuroprotection, behavioral improvement, and so on. One of the most recognized molecular mechanisms of tPBM is that cytochrome c oxidase (CCO) may dissociate inhibitory nitric oxide (NO) after absorbing photons, thereby enhancing mitochondrial activity and promoting ATP biosynthesis.¹⁸ Because brain disorders are closely related to mitochondrial activity, tPBM may have beneficial effects on various brain diseases.¹⁹ During the past two decades, a tremendous number of pre-clinical experiments with animals and clinical trials with humans have demonstrated beneficial effects. Clinical trials with humans include, in particular, the treatment of ischemic stroke,²⁰⁻²⁴ Alzheimer's disease (AD),²⁵⁻²⁸ Parkinson's disease (PD),²⁹⁻³⁴ traumatic brain injury (TBI),³⁵⁻⁴⁰ depression,⁴¹⁻⁴⁴ aging,⁴⁵⁻⁵⁰ etc. In these studies, the parameters of the light used are extremely sophisticated and deserve to be deliberated. For wavelength, researchers prefer using 808 nm-light [Figs. 1(a) and 1(b)], which has a strong absorption peak of CCO with an excellent penetration depth.¹⁴² Furthermore, there also exist extensive studies that have applied other wavelengths (e.g., 610, 1070, 1267 nm) of light for brain disease treatment for specific biological targets including but not limited to mitochondrial stimulation.^{51,74,75,143} In particular. compared with traditional laser therapy (630 to 900 nm), the light in the NIR-II region (1000 to 1700 nm) has much less scattering and thus can penetrate deeper into the brain despite its slighter stronger absorption.^{144–146} Researchers have demonstrated that the light in the NIR-II region presents the absence of carcinogenic or mutagenic properties.¹⁴³ The utilization of 1267 nm tPBM represents a burgeoning strategy in the therapeutic landscape of brain diseases, attributable to its recently unveiled unique capacity to stimulate the brain waste removal system (BWRS), thus contributing to neuroprotection of the CNS.¹⁴⁷ It has been reported that such a wavelength can activate the generation of singlet oxygen in biological tissues,^{148,149} which can lead to vascular responses and intense elimination of toxins and unnecessary molecules from the brain.^{150–152} In addition, pulsed wave (PW) light has been found to penetrate deeper into tissues than continuous wave (CW) light with the same average power, as supported by theories and ex vivo experiments.^{153–156} Moreover, PW light with a specific frequency may activate specific ion channels (e.g. transient receptor potential channel) to trigger a series of beneficial biological responses.⁷⁵ For the light source, whether a coherent monochromatic laser is superior to incoherent light emitting diodes (LEDs) is an ongoing debate, with some researchers affirming^{157,158} but some denying.¹⁵⁹ Although lasers were preferred at the early stage, especially for preclinical trials, LEDs are widely designed as helmet-type devices for clinical trials due to their low price and easy assembly [Figs. 1(c) and 1(d)]. Figure 2 shows representative tPBM devices for treating various brain diseases, including PD,³¹ TBI,³⁶ depression,⁴³ and androgenetic alopecia.¹⁶⁰

In this review, we shed light on the tPBM-mediated mechanisms of therapy for brain diseases according to the emerging trends in tPBM and new discoveries. We also review the

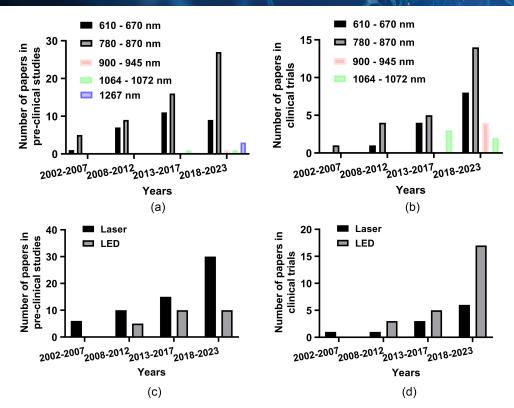


Fig. 1 Number of published articles using different tPBM wavelengths and light sources for brain disease treatment in pre-clinical studies and clinical trials from 2002 to 2023. (a) The number of published articles using different tPBM wavelengths in pre-clinical studies. (b) The number of published articles using different tPBM wavelengths in clinical trials. (c) The number of published articles using different tPBM sources in pre-clinical studies. (d) The number of published articles using different tPBM sources in pre-clinical studies. (d) The number of published articles using different tPBM sources in pre-clinical studies. (d) The number of published articles using different tPBM sources in clinical trials. Notes: the data are summarized from Refs. 20–23 and 51–70, (ischemic stroke), 71–73 (HI), 74 (intracerebral hemorrhage), 25–28, 75–83 (AD), 29–34, 84–96 (PD), 97–99 (multiple sclerosis), 35–40, 100–116 (TBI), 117 (possible chronic traumatic encephalopathy), 41–44, 118–126 (depression), 45–50, 127–136 (aging), and 137–141 (epilepsy).

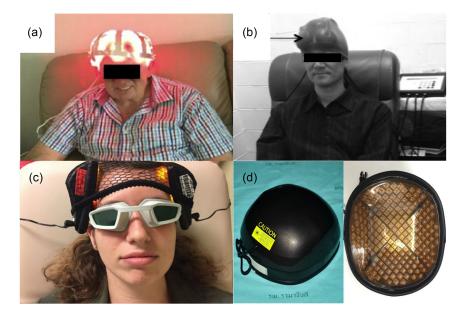


Fig. 2 Examples of tPBM helmet devices for clinical treatment of brain diseases: (a) PD,³¹ (b) TBI,³⁶ (c) depression,⁴³ and (d) androgenetic alopecia.¹⁶⁰

2 Main Mechanisms of tPBM

2.1 Molecular and Cellular Mechanisms of tPBM

The molecular and cellular mechanisms of tPBM are very complex (Fig. 3), with the enhancement of mitochondrial activity recognized as one of the most widely studied and crucial mechanisms.^{161,162} Mitochondrial CCO is the terminal enzyme in the mitochondrial respiratory chain that contributes an increase in metabolic and energetic activity of cells via more oxygen consumption. Both *in vitro*¹⁶³ and *in vivo*^{47,164–166} studies have revealed that red and NIR light (e.g., 660, 808, 850, and 1064 nm) can significantly increase the CCO activity and expression level. After boosting CCO, the mitochondrial membrane potential (MMP) is increased, more oxygen is consumed, more glucose is metabolized, and more ATP is produced.¹⁶⁷ Furthermore, a recent study proposed that 1267 nm laser-generated singlet oxygen can work as an activator of mitochondrial respiration and ATP production in brain cells.¹⁶⁸ The above evidence demonstrates that tPBM can increase cell respiration, boost brain energy metabolic capacity, enhance brain electrophysiological oscillation (alpha and beta bands) strength, and improve cerebral oxygenation,^{169–172} which would constitute an adaptation with major neuroprotective implications after brain disease.

Reactive oxygen species (ROS) is an umbrella term for an array of derivatives of molecular oxygen. It was reported that the release of low-level mitochondrial ROS is involved in regulating transcription factors and signaling mediators, potentially leading to beneficial effects.¹⁷³ Meanwhile, downregulation of excessive ROS may also help inhibit oxidative stress and neuro-inflammation, thereby protecting the neuronal mitochondria. Currently, the regulatory effects of parenchymal border macrophages (PBM) on ROS are still controversial. Some studies have shown that PBM enhanced the ROS production by activating the superoxide converting system,^{174,175} whereas other studies have suggested the opposite.¹⁷⁶ A recent study found that PBM increased ROS levels in normal neurons but reduced it in oxidatively stressed neurons.¹⁷⁷

The brain diseases are typically accompanied by oxidative stress damage. Therefore, reducing ROS levels may provide a potential possibility for brain disorder treatment. Salehpour et al.

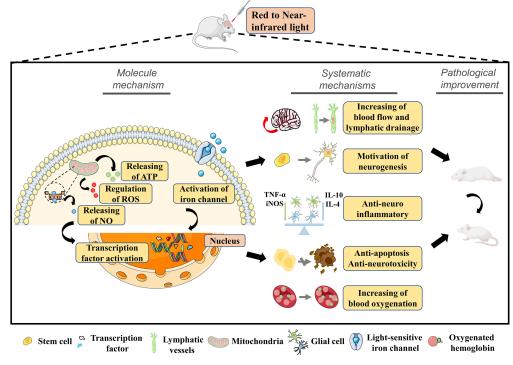


Fig. 3 Molecular mechanisms and neurobiology effects of tPBM.

found that tPBM (810 nm) reduced the mitochondrial ROS levels and mitigated learning and memory impairments in aging¹²⁷ and in ischemic stroke¹²⁸ mouse models. Zhang's team found that tPBM (808 nm) significantly inhibited the oxidative damage induced by the increase of superoxide anion after beta-amyloid ($A\beta$) intraventricular injection and significantly enhanced the total antioxidant capacity.⁷⁶ In addition, they observed that tPBM (808 nm) increased the total antioxidant capacity in a TGF344 AD transgenic rat model and reduced the level of oxidative stress products (e.g., lipid peroxide malondialdehyde).⁷⁷ Similar beneficial effects mediated by tPBM have also been shown in mice with sleep deprivation and depression.^{118,119}

Neuroinflammation is one of the crucial pathophysiological conditions associated with various brain disorders. tPBM can decrease the expression of pro-inflammatory cytokines via inhibition of NF- κ B signaling pathways.¹⁷⁸ It can also directly alter the phenotype of glial cell and then regulate the release of inflammatory cytokines. Specifically, tPBM reduces the number of cortical pro-inflammatory phenotype (M1) microglia in ischemic stroke and brain trauma mouse models.^{52,100} In addition, some studies have demonstrated that tPBM promotes the polarization of cortical and hippocampal microglia from M1 to an anti-inflammatory phenotype (M2) in AD⁷⁷ and neonatal hypoxic-ischemic (HI) rat models.⁷¹ Furthermore, tPBM can improve the lymph flow in the meningeal lymphatic vessels (MLVs), thereby promoting removal of pro-inflammatory cytokines from the brain.¹⁷⁹

In addition to the mechanisms mentioned above, other molecular mechanisms such as gating of the channelrhodopsins and the regulation of the transcription factor hypoxia-inducible factor (HIF-1 α) may also contribute to the beneficial effects of tPBM.¹⁸⁰ In summary, the interaction between photons and tissues is highly complex, and the mechanism of tPBM is not yet well understood. Further exploration of the mechanism of tPBM will have important implications for clinical treatment of brain diseases.

2.2 Systemic Mechanisms of tPBM

Recently, new effects of tPBM on the BWRS have been discovered via tPBM-induced stimulation of MLVs.^{78,79,147,181–185} In vivo and in vitro experiments have shown that PBM induces relaxation of lymphatic endothelium of both the MLVs and the mesenteric lymphatic vessels (LVs).¹⁸⁴ PBM also increases the permeability of lymphatic endothelium due to a decrease in the expression of the tight junction (TJ) proteins.¹⁸⁴ These effects of PBM might be related to a PBM-related stimulation of the NO synthesis in the lymphatic endothelium.^{17,74,186} The NO causes the relaxation of the blood and LVs through soluble guanylate cyclase and protein kinase G, leading to reducing of the Ca^{2+} intracellular levels and blocking phosphorylation of the myosin light-chain kinase.¹⁸⁷ PBM causes vascular relaxation also via an increase of the activity of endothelial NO synthase (eNOS).^{17,53} The eNOS induces the NO production in the lymphatic endothelium that promotes an increase in lymphatic flow and removal of wastes and toxins (e.g., $A\beta$) from the brain and other tissues.^{74,78,188} Meanwhile, the increase in relaxation of MLVs mediated by tPBM also opens promising perspectives for lymphatic delivery of nanocarriers and drugs to the brain pathology bypassing the BBB. It was reported that a 1267 nm laser enhances the lymphatic transport of liposomes from the deep cervical lymph node (dcLN) to brain parenchyma and promotes the clearance of these nanocarriers from the subarachnoid space, offering a novel potential strategy for treating brain disease (Fig. 4).¹⁸² NO also controls lymphatic contractility, which is important for the movement of cells and molecules in the LVs.¹⁸⁹ A hypothesis posits that a PBM-mediated release of NO could improve the lymphatic contractility, which might be another possible mechanism underlying PBM-activation of clearance of wastes and toxins from the brain (Fig. 4).74,180,182,183,190 Li et al. proposed that tPBM (1267 nm) reduced the mortality of an intraventricular hemorrhage (IVH) mouse model by promoting red blood cells (RBCs) transport from the ventricle to dcLNs. Significantly, they revealed that this beneficial effect disappeared when NO was blocked.⁷⁴ In addition, it has been reported that sodium nitroprusside can increase the sensitivity of LVs to PBM,¹⁹¹ providing a potential strategy to further enhance the effects of tPBM treatment.

In additional to facilitating lymphatic drainage, the release of NO during tPBM could also increase cerebral blood flow¹⁹² and might contribute to the enhancement of cerebral endogenic and myogenic functional connectivity (FC).^{171,193–195}

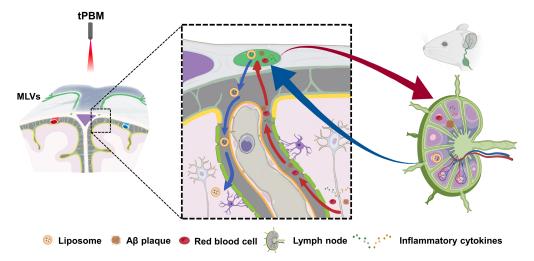


Fig. 4 Illustration of new tPBM strategy for brain disease treatment: red and blue arrows represent MLVs-mediated toxins evacuation pathway and liposome-loaded drug delivery pathway, respectively. Some items were created with BioRender.

Except for tPBM-mediated NO regulation, it has been demonstrated that tPBM can attenuate cerebral $A\beta$ burden through the activation of the cAMP-dependent protein kinase signaling pathway, mediated by CCO, as well as via the stimulation of microglia and angiogenesis.^{75,80,196} In addition, Tao et al. reported that PBM (1070 nm) reduces the $A\beta$ levels in the brain via stimulating and recruiting microglia to the $A\beta$ burden⁷⁵ (Fig. 5) and increasing cerebral vessel density.^{75,196} Yue et al. found that tPBM (630 nm) improves brain drainage leading to the $A\beta$ removal in the APP/PS1 mouse model of AD.⁸¹

The release of the brain-derived neurotrophic factor (BDNF) may also contribute to PBMmediated stimulation of $A\beta$ elimination from the brain. Experiments *in vitro* indicated that PBM (632 nm) increases BDNF levels by activating the extracellular signal-regulated kinase/cyclic AMP-responsive-element-binding protein (CREB) pathway, which ameliorates $A\beta$ -induced neuronal damage and dendritic atrophy.¹⁹⁷ In addition, it has been demonstrated that tPBM (810 nm) significantly increases the BDNF level in the hippocampus of mice with brain trauma,¹⁰⁰ depression,¹²⁰ and ischemic stroke,¹⁹⁸ which may be beneficial to neurogenesis and synaptogenesis.¹⁹⁹

PBM with a light wavelength of 1267 nm can stimulate the generation of singlet oxygen directly in cultured cells and biological tissues without photosensitizers.^{148,149,200,201} Stanley et al. reported a new mechanism of singlet oxygen to regulate the endothelial relaxation via the control of the vascular tone.^{150,152} The singlet oxygen induces oxidation of the amino acid tryptophan in the mammalian tissues that leads cell production of metabolite, such as N-formylkynurenine with

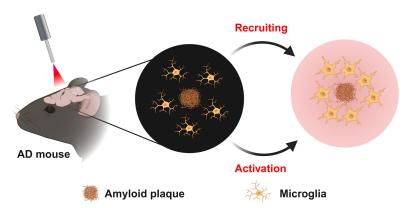


Fig. 5 Illustration of a mechanism for tPBM treatment of AD: stimulating and recruiting microglia to the $A\beta$ deposition.

2.3 New Strategy in the Study of tPBM-Mechanisms

Glioblastoma (GBM) is the most common and aggressive form of brain cancer and is one of the deadliest brain tumors.²⁰² Recently, it was discovered that GBM is characterized by reducing functions of the MLVs, which play an important role in the regulation of brain tumor drainage and immunity.^{203–206} These findings provide new strategies in the stimulation of an efficient immune response against glioma.^{203,207} Hu et al. revealed significant changes in the gene controlling the MLVs remodeling, fluid drainage, and inflammatory and immunological responses in mice with GBT.²⁰³ They discovered that mice with GBT and overexpression of the vascular endothelial growth factor C (VEGF-C) demonstrate a better response to anti-tumor therapy, such as the combination of anti-programmed death-1 protein (PD1) and cytotoxic T-lymphocyteassociated protein 4 (CTLA-4) blockade. It is interesting that the blockade of the MLVs abolished this effect.²⁰³ Indeed, mice with GBT treated with blockers of lymphatic proteins, such as chemokine (C-C motif) ligand 21 (CCL21) and C chemokine receptor type 7 (CCR7), did not show the improvement of anti-tumor therapy.²⁰³ Authors concluded that VEGF-C potentiates checkpoint therapy via the CCL21/CCR7 pathway. Song et al. also reported that VEGF receptor-3 (VEGFR3) enhances immune surveillance from GBM and improves the effectiveness of anti-tumor therapy with checkpoint inhibitors.²⁰⁸ Both Hu et al.²⁰³ and Song et al.²⁰⁸ documented an important role of the MLVs in the regulation of brain tumor immunity in mice. Because the tPBM-effects on the MLVs have already been documented,^{17,74} these pioneering results open new perspectives for the study of the role of tPBM in the stimulation of the CCL21/CCR7 pathway. This mechanism underlies the tPBM-induced effects on the MLVs and presents an alternative strategy for GBM treatment.

PBMs are a newly defined cell population. They reside in the leptomeninges and perivascular spaces along the vasculature and are responsible for regulating the cerebrospinal fluid flow dynamic. Researchers found that activating PBMs via intracisternal injection of the macrophage colony-stimulating factor could restore CSF dynamics in aging mice,²⁰⁹ which indicates that PBM activation via tPBM may help alleviate brain clearance deficits associated with aging and AD.

3 tPBM for Brain Diseases

3.1 tPBM for Ischemic Stroke

Ischemic stroke is a sudden cerebrovascular disease with high mortality. Some studies have proved that the BBB damage caused by ischemic stroke could increase the permeability of blood vessels, resulting in blood cells and neurotoxins entering the brain parenchyma and causing brain damage. It may activate inflammatory signals, trigger biochemical and molecular events, and further aggravate brain damage.²¹⁰ In addition, ischemic stroke may alter the mitochondrial morphology and permeability, leading to the decrease of MMP and ATP levels, thereby inducing cell damage and apoptosis.²¹¹ Tissue plasminogen activator (TPA) is the only therapeutic approved by the U.S. Food and Drug Administration (FDA), but it must be received within 4.5 h after stroke for the effectiveness,²¹² and TPA may cause a more serious cerebral hemorrhage.

tPBM has the effects of mitochondrial function improvement and anti-inflammation, which is a potential therapeutic approach for ischemic stroke (see Table S1 in the Supplementary Material). In pre-clinical studies, Lee et al. found that transcranial LED light (610 nm) significantly reduced the ischemic infarct size, improved the neurological function score, inhibited neuroinflammation, and attenuated neuronal apoptosis in the photothrombotic mouse model.⁵⁴ In addition, they also observed that applying the same tPBM protocol within 4 h after stroke improved long-term functional recovery and promoted neurogenesis and angiogenesis in a cerebral ischemic mouse model. This suggests that the therapeutic time window of certain tPBM was at least 4 h.⁵⁵ Importantly, they also found that pretreatment with tPBM prevented the BBB damage and the blood flow decrease caused by ischemic brain injury.^{51,53} These beneficial effects were associated with the increase of expression of the TJ protein and the NO release in the blood endothelium. Recently, Kim et al. developed an implantable multi-LED (630 nm) array, which could prevent tissue and functional impairment, and attenuated the cognitive decline in an acute ischemic stroke mouse model.⁵² In addition, tPBM with wavelengths of 660,⁵⁶ 780,⁵⁷ 808,⁵⁸⁻⁶⁰ and 904⁶¹ nm also has beneficial effects of neuroprotection, anti-inflammation, and cognitive improvement in ischemic stroke rat models (including the middle cerebral artery occlusion model and the photothrombotic model). Oron et al. found that tPBM in the CW mode could better improve the neurological score and promote the neurogenesis in ischemic stroke mice than tPBM in the pulsed irradiation.^{58,62} Moreover, Lapchak et al. applied tPBM (808 nm) to the rabbit small clot electromagnetic stroke model to further verify the effectiveness. Specifically, they observed that applying tPBM with a power density of 7.5 mW/cm² within 3 h post-embolization significantly improved clinical rating scores. In addition, the therapeutic time window could be extended to 6 h by increasing the laser power density to 25 mW/cm^{2,63} Similar dose-dependent beneficial effects were also observed by Huisa's team.⁶⁴ However, excessive power density may also lead to the plateau effect⁶⁵ or even the biphasic effect.⁶⁶ In addition, Lapchak et al. found that pulsed light (100 Hz) instead of continuous light could completely inhibit the decrease of ATP levels in the embolic cortex.^{65,67} Furthermore, it was reported that tPBM combined with TPA could better improve the behavior performance and mitigate the ATP decrease in the ischemic cortex.^{68,69}

The clinical results of the NeuroThera Effectiveness and Safety Trial-1 (NEST-1) showed that tPBM (808 nm) within 24 h after stroke is safe and effective on clinical ischemic stroke treatment.²⁰ However, the NEST-2 trial results showed that tPBM did not have a significant beneficial effect when the patient count and age distribution were increased.²¹ In addition, NEST-3 revealed that tPBM has no measurable neuroprotective effects in stroke patients.⁷⁰ The above earlier studies may have had better results if more than one tPBM treatment had been applied and if tPBM had been applied to only the same side of the head, such as where the stroke had occurred. Naeser et al. reported that the naming ability in left-hemisphere (LH) stroke patients with lasting language problems (aphasia) was significant improved following 18 tPBM treatments applied to only one side of the head/scalp where the stroke had occurred. However, no improvement occurred when tPBM was applied to both sides of the head.²² In addition, a recent study demonstrated that the combination of tPBM and speech-language therapy resulted in greater improvements in speech-language skills post-stroke compared with speech-language therapy alone.²³

3.2 tPBM for Hypoxic-Ischemic

An HI brain injury is a common clinical birth complication that has high mortality and disability rates.²¹³ Currently, the only licensed therapeutic approach in clinic is hypothermia. However, it not only has a very narrow therapeutic time window but also has limited efficacy and usually comes with potential adverse cardiovascular effects.²¹⁴ Therefore, there is an urgent need for a more practical and effective therapy for HI. It was reported that HI mainly damages mitochondria,⁷¹ causing mitochondrial structure (e.g., mitochondrial fragmentation) and function (e.g., reducing ATP production) impairment, thereby inducing oxidative stress and neuroinflammation. Therefore, protecting mitochondria from damage or enhancing mitochondrial function is a potential strategy for HI treatment.

Given that NIR light treatment has the potential to improve the mitochondrial function, Zhang's laboratory explored whether tPBM (808 nm) has beneficial effects on neonatal HI (see Table S2 in the Supplementary Material). First, they found that applying tPBM immediately after HI for seven consecutive days contributed to robust neuroprotection effects by mitigating mitochondrial dysfunction, oxidative stress, and final neuronal apoptosis in rats.⁷² Second, they found that tPBM administered once 6 h before HI induction in postnatal 10 rats also mitigated brain damage.⁷³ They also reported that applying tPBM three times per week to the abdomen of pregnant rats from gestation day (GD) 1 to GD 21 also exerted the neuroprotective effects against hypoxic ischemia in rat pups.⁷¹ In addition, Gerace et al. developed a dual-wavelength (808 and 904 nm) NIR laser source device that could attenuate the neurotoxicity of oxygen and glucose

deprivation in hippocampal slices by attenuating inflammation.²¹⁵ The above findings clearly show that tPBM might be a promising technology for therapy of HI in neonates. Further confirmation of animal data in clinical investigations will open a new niche of tPBM application in neonatology.

3.3 tPBM for Intracerebral Hemorrhage

IVH is defined as bleeding within the brain's ventricular system, a condition associated with high rates of morbidity and mortality.²¹⁶ Surgery and fibrinolysis in combination with extraventricular drainage is the conventional therapy of IVH. However, it has not made a significant impact on the natural history of IVH. Thus, new therapeutic approaches need to be urgently found to mitigate hematoma expansion and improve the drainage system of the brain.²¹⁷ Li et al. demonstrated that an NIR tPBM (1267 nm) accelerates the RBCs evacuation from the ventricles to the dcLNs via the MLVs, leading to improvement of the neurological status in mice and newborn rat pups and their recovery after IVH (see Table S3 in the Supplementary Material).⁷⁴

Li et al. optimized the tPBM illumination parameters for hemorrhagic stroke using the Monte Carlo method to stimulate photon propagation within the visible Chinese human head at a different level of intracerebral hemorrhage with varied parameters of light beams. They found that the Gaussian beam with a similar or larger size as the hemorrhagic region had the best therapeutic outcomes, whereas the top-hat beam performed better when the hemorrhagic region was much bigger than the beam size.^{142,218}

3.4 tPBM for Alzheimer's Disease

AD is a neurodegenerative disease that dramatically reduces the quality of life and ultimately leads to death.²¹⁹ However, there is a lack of effective therapeutic strategies for AD in clinical practice. The amyloid cascade hypothesis proposes that the deposition of the A β peptide in the brain is a central event in disease pathology, which has long been the primary focus to develop therapeutic approaches that may slow or delay the progression of AD.²²⁰

Currently, there is compelling evidence suggesting that tPBM is capable of decreasing the Aβ burden and ameliorating cognitive and memory impairments in the AD animal models in various pathways (see Table S4 in the Supplementary Material). The latest trends suggest that tPBM (1267 nm),⁷⁹ especially during deep sleep,⁷⁸ stimulates lymphatic removal of A β from the mouse brain. A significant reduction of A β plaques in the hippocampus CA1 region was associated with improved recognition memory and cognitive status in mice (injection model of AD). This can be explained because night tPBM has a synergistic effect on the natural activation of mechanisms underlying night lymphatic drainage of brain parenchyma. Moreover, Xing's lab proposed that tPBM (633 nm) shifted the amyloid precursor protein (APP) processing toward the nonamyloidogenic pathway by activating Sirtuin 1 via the cyclic adenosine monophosphate/ protein kinase pathway, thereby improving memory and cognitive ability in an AD mouse model.⁸⁰ They also showed that tPBM (633nm) could inhibit the activity of c-Jun N-terminal kinase 3 (a signal molecule related to neurodegeneration); this may be of therapeutic utility in the treatment of AD.⁸² Recently, Yue et al. provided new insight for AD treatment that showed that the use of a 630 nm laser in treatment could reverse A β -obstructed interstitial fluid flow and ameliorate memory decline in APP/PS1 mice.⁸¹ Grillo et al. observed 1072 nm laser treatment upregulated some stress response proteins in the AD mouse brain, known to reduce both $A\beta$ aggregation and neuronal apoptosis.⁸³ Tao et al. demonstrated that LED treatment at 1070 nm attenuated the A β burden by modulating the microglia phagocytosis capacity and promoting angiogenesis in AD mice.⁷⁵ Zhang's lab demonstrated that tPBM (808 nm) could not only mitigate A β -induced pathology (e.g., mitochondrial dysfunction, neuronal apoptosis, and tau pathology)⁷⁶ but also prevent or slow the progression of AD in a rat model.⁷⁷

In clinical trials, Berman et al. found that the application of tPBM (1060 to 1080 nm) for 28 consecutive days improved cognitive and memory ability in AD patients.²⁸ In addition, three more recent tPBM clinical studies on mild to moderately severe dementia cases have been published; these included subjects who likely had a progressive neurodegenerative disease, e.g., AD, and tPBM was observed to improve cognition, mood, and sleep.^{25–27} Cases with MMSE entry scores of 10 to 24/30 and treated for 12 weeks showed significant cognitive improvements soon after the final tPBM treatment. However, 1 month later, there was a decline

in their scores.²⁵ Collectively, the above research with animal studies and early results from human clinical studies have put forward some tPBM mechanisms for mitigating AD, thus shedding light on a possible new treatment.

3.5 tPBM for Parkinson's Disease

PD is a progressive neurodegenerative disorder, with more than 6 million patients worldwide. It is characterized by the loss of dopaminergic neurons in the substantia nigra and the presence of lewy bodies in the midbrain.²²¹ Currently, the only available therapeutic approach for PD management is mainly based on exogenous dopaminergic supplements, such as levodopa. Such a treatment cannot modify the disease course or slow the underlying neurodegeneration associated with the multifactorial characteristics of PD.²²² Some accepted points of view are that mitochondrial dysfunction, oxidative stress, and protein mishandling have a central role in PD pathogenesis.²²³ Furthermore, it has been reported that improving mitochondrial structure and function, thereby mitigating oxidative stress, could have beneficial effects for PD management.²²⁴ Consequently, tPBM has emerged as a potential therapeutic approach for PD. Based on this hypothesis, researchers have performed pre-clinical studies and clinical trials to explore whether tPBM could effectively treat PD. Some early results are provided in Table S5 in the Supplementary Material.

In pre-clinical trials, numerous studies demonstrated that tPBM (670 and 810 nm) effectively alleviated PD pathology in a 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)induced PD mouse model. This included the prevention of the loss of tyrosine hydroxylasepositive (TH⁺) cell and cerebrovascular leakage in the substantia nigra pars compacta (SNc) region.⁸⁴⁻⁹⁰ In addition, the neuroprotective effects of tPBM were also observed in lipopolysaccharide (LPS)-induced,⁹¹ 6-hydroxydopamine (6-OHDA)-induced,⁹² AAV2/6 virus-induced,⁹³ and transgenic PD rat models⁹⁴ and an MPTP-induced macaque monkey model.⁹⁵ It should be noted that various factors may affect light penetration from the skull into the brain, thus affecting the therapeutic effects. Salgado et al. found that transcranial laser treatment, rather than LED, significantly reduced the level of serum pro-inflammatory cytokines.⁹² Mitrofanis' group showed that the neuroprotective benefits of tPBM (670 nm) were more effective in the BALB/C strain than in the C57BL/6 strain. This could be attributed to deeper photon penetration, far into the brain from the skin in the BALB/C mice.89,225 They also discovered that the neuroprotective effects of tPBM existed in the SNc region but not in the periaqueductal grey matter (PaG) and zona incerta-hypothalamus (ZI-Hyp) regions. They attributed this to the fact that cell characteristics and photon penetration differ in different brain regions.⁸⁴ Interestingly, Johnstone et al. observed that irradiation targeted to the dorsum and hindlimb also had neuroprotective effects, although not as effective as transcranial irradiation. This remote effect for PD treatment may be mediated by the regulation of variable signaling paths, such as stem cell-related C-X-C chemokine receptor type 4 signaling and oxidative stress response pathways.^{96,226}

In clinical trials, McGee et al. found that tPBM (635 nm plus 810 nm LEDs) could safely and meaningfully improve individual motor signs of PD.³³ Hamilton et al. developed a helmet device with several LEDs with wavelengths of 670, 810, and 850 nm and reported improvement in PD progression.³¹ In fact, owing to the exponential attenuation of light traveling through skull and brain tissues, only a very limited number of neurons can absorb sufficient photons for positive effects. Therefore, various PBM strategies were developed for clinical PD treatment. Liebert et al. stressed the importance and demonstrated the effectiveness of treating the abdomen and the brain with PBM (gut/brain axis) in PD.³⁰ Other light delivery strategies, such as neck and intranasal PBM, were also used in combination with tPBM for clinical PD treatment and reported effectiveness.^{29,34} Hong et al. found that tPBM (940 nm) targeting the brainstem combined with molecular hydrogen treatment significantly reduced the Unified Parkinson Disease Rating Scale scores in PD patients. Such an improvement still existed in the follow-up period of 1 week after treatment.³² In summary, the above encouraging results of pre-clinical studies and clinical trials show the great potential of tPBM in PD treatment.

3.6 tPBM for Multiple Sclerosis

MS is one of the most common chronic inflammatory, demyelinating, and neurodegenerative diseases of the CNS in adults; it affects two to three million people worldwide.²²⁷ One of the

the regulation of inflammation and the promotion of neurogenesis.⁹⁷

In pre-clinical studies, Duarte et al. found that tPBM improved motor performance in mice with demyelination. This improvement may be associated with the attenuation of demyelination, proliferation of oligodendrocyte precursor cells, and inhibition of neuroinflammation.⁹⁸ In clinical trials, Silva et al. found that tPBM effectively upregulated the expression level of interleukin-10 in MS patients.⁹⁷ They also found that tPBM improved the fatigue status of MS patients, despite no statistical significance in the modified fatigue impact scale measurement.⁹⁹ These studies (see Table S6 in the Supplementary Material) have suggested evidence for support of tPBM as a possible treatment for MS. Future investigations are likely to provide positive results.

3.7 tPBM for Traumatic Brain Injury

TBI is defined as an alteration in brain function or other evidence of brain pathology caused by an external trauma.²²⁹ The pathophysiology of TBI is highly heterogeneous and complex, including adverse signaling pathways activation, inflammation, oxidative stress, mitochondrial dysfunction, and excitotoxic damage. The combination of cellular and physiological disturbances increases the infarct size, neurological decline, and cognitive impairment. TBI occurs more than 50 million times annually worldwide, posing a significant burden on socio-economic and health-care systems.²³⁰ Currently, due to the heterogeneity of TBI and limited understanding of potential pathophysiological mechanisms, there are no standardized methods or drug treatments.²³¹ This has led to interest in new treatment approaches such as tPBM. Since the 21st century, many research teams have begun to explore the potential of tPBM as a treatment for acute or chronic TBI. There have been many positive results (see Table S7 in the Supplementary Material).

Hamblin's team found that one transcranial, red (660 nm), or NIR (810 nm) laser treatment performed at 4 h after TBI significantly improved the neurological severity score (NSS) and decreased the brain lesion volume in moderate-to-severe TBI mouse models.^{101,102} They also reported that tPBM, when applied during three consecutive days post-TBI, was more effective than only one irradiation, in improving the motor and memory ability in the TBI mouse model.¹⁰³ This reduced the degeneration and apoptosis of neurons in the injured region,^{103,104} promoting the neurogenesis, synapse formation, and expression level of BDNF in the dentate gyrus (DG) of the hippocampus and in the subventricular zone.^{104,105} However, it was also reported that an excessive number of tPBM treatments in mice with TBI could temporarily inhibit the process of brain repair, suggesting that it is important to choose the optimal protocol of tPBM for TBI.¹⁰⁶ Shemesh et al. found that tPBM (810 nm) regulated the hemodynamics, with reduced cell death and stimulation of neurogenesis.¹⁰⁷ Wu's lab found that tPBM (810 nm), combined with energy metabolism regulators (e.g., lactic acid or pyruvate), more effectively increased ATP levels in the impact cortex and reduced neuronal damage and neuroinflammation caused by the TBI.^{108,109} Micci's team found that tPBM combined with ultrasound (optoacoustic) treatment effectively mitigated sympathetic dysfunction, neuroinflammation, and dysregulation of neurogenesis in the blast brain injury mouse model.^{100,110} Similarly, neuroprotective effects of tPBM for TBI in pre-clinical studies were observed in the laboratories of Whalan,^{111,112} Marques,¹¹³ Oron,^{114,115} and Zhang.¹¹⁶

Some of the above studies compared therapeutic effects from different light parameters. For various wavelengths, Reinhart et al. found that tPBM at 660 and 810 nm, but not at 730 or 980 nm, had neuroprotective effects.¹⁰² This can be due to weak light absorption by CCO of 730 and 980 nm.¹⁰² Regarding the delivery mode, Oron et al. found that the 100-Hz pulsed laser improved the NSS better than CW and the 600-Hz pulsed laser in a TBI mouse model. The authors speculated that this phenomenon may be related to the resonance effect between 100-Hz PW laser and brain waves (such as alpha and theta waves).¹¹⁵ Also, Ando et al. proposed that a 10-Hz pulsed laser has more neuroprotective and cognitive improvement effects than a 100-Hz PW laser. This may be induced by a positive resonance between the 10-Hz PW laser and the electrical activity of neurons in the hippocampus.¹⁰¹ Both Abookasis and Whalen's team

In clinical trials, Naeser et al. found that tPBM (at 633 and 870 nm) treatment significantly improved the cognitive performance in mild TBI patients.³⁶ The team also reported that tPBM could enhance sleep duration by an average of 1 h in chronic TBI cases,^{37,38} for which poor sleep is a common complaint. Longo et al. studied acute, hospitalized, moderate TBI cases with magnetic resonance imaging (MRI) and reported significant differences between MRI-derived diffusion parameters in white matter tracts between sham versus real LED-treated groups, demonstrating safety and neuro-reactivity for real tPBM in this population.³⁹ Nawashiro et al. used single-photon emission computed tomography brain scans, after 73 days of LED (850 nm) tPBM, applied twice a day to the left and the right forehead areas and reported that the regional CBF (rCBF) increased by 20% in the left anterior frontal lobe in a severe TBI patient who was in a persistent vegetative state.⁴⁰ This was associated with some new arm movement. In addition, Chao et al. reported that after combining intranasal plus transcranial PBM (810 nm) treatments for 8 weeks, there was increased brain volume, improved FC, increased cerebral perfusion, and improved neuropsychological test scores in an athlete, age 23, who had had six concussions in 5.5 years.³⁵

In conclusion, tPBM for TBI mitigates the death of brain neurons, decreases neuroinflammation, and improves the self-repair ability of the brain by stimulating synapses formation and proliferation of nerve cells, demonstrating high potential of tPBM for the clinical treatment of TBI.

3.8 tPBM for Possible Chronic Traumatic Encephalopathy

CTE is a progressive neurodegenerative disease present in athletes who have sustained repetitive head impacts. At post-mortem, hyper-phosphorylated tau deposits (p-tau) are present in the deep sulcal areas, which is unique to CTE. It differs from AD, in which A β and another form of tau deposits are present in different areas of the brain. Symptoms of CTE include behavioral and mood changes, memory loss, cognitive impairment, and dementia.²³² However, there are no known current treatments for CTE.²³³ Recently, Naeser et al. reported the new use of tPBM to treat possible CTE.¹¹⁷ This study showed significant improvements in cognition and behavior/mood for four ex-football players after 18 tPBM treatments (see Table S8 in the Supplementary Material). At 2 months after this first in-office, tPBM treatment series was completed, two of these ex-football players regressed. Then, home tPBM treatments were selfapplied to only cortical node areas of the default mode network (12 weeks). Again, significant improvements returned. Increased FC for the salience network, post-tPBM, was present. Also, increased n-acetyl-aspartate (NAA), reflecting increased oxygen consumption in the mitochondria of neurons, was present in the anterior cingulate cortex, parallel to less pain and PTSD, and lasted for 12 weeks post-tPBM. Ongoing, tPBM treatments that can be safely applied at home may be necessary to maintain gains in cases with progressive neurodegenerative disease.

3.9 tPBM for Depression

Depressive disorder is one of the most prevalent and debilitating forms of psychopathology;²³⁴ it manifests by lack of energy, depressed mood, low executive ability, and poor concentration. Epidemiological surveys indicate that depressive disorder affects more than 16% of the worldwide population. This increases the prevalence of medical illnesses, such as type 2 diabetes, cardiovascular diseases, and autoimmune diseases, and it even advances biological aging.²³⁵ At present, pharmaceutical intervention is the most mainstream therapeutic approach for depression, but it has a low effective rate and even some opposite reactions. Electroconvulsive therapy (ECT), TMS, and vague nerve stimulation are FDA-approved neuromodulation strategies for the treatment of depression. However, complex procedures, such as anesthesia for ECT, as well as expensive costs limit their clinical application.²³⁶ Therefore, there is urgent need for a safe, effective, and convenient treatment strategy for depression that may be well tolerated and potentially used by patients at home. It has been reported that mitochondrial dysfunction, cerebral energy metabolism impairment, and oxidative stress may play a significant role in the development of depression.²³⁷ tPBM has been reported to modulate a variety of biological processes, including anti-oxidation, anti-inflammation, neuro-enhancement, ATP synthesis, etc.²³⁸ This suggests that

tPBM may be an attractive, new treatment for depression. Currently, extensive pre-clinical studies and clinical trials have demonstrated that tPBM indeed has beneficial effects on depression (see Table S9 in the Supplementary Material).

Salehpour et al. found that tPBM (810 nm) significantly improved the neurological status in sleep-deprived and, in restraint, stress-induced depression mouse models. The authors proposed that reversing oxidative stress, neuroinflammation, and neuronal apoptosis in the prefrontal cortex (PFC) and hippocampus might be a target for the beneficial effects of tPBM in these models.^{118,119} Xu et al. found that tPBM (808 nm) elevated PFC ATP levels and mitochondrial complex IV activity in space restriction and abelson helper integration site-1 (Ahi1) knockout (KO) depression mouse models.¹²¹ Farazi found that tPBM (810 nm) attenuated the decrease of BDNF, tropomyosin receptor kinase B (TrkB), and phospho-CREB/CREB in the hippocampus and down-regulated the serum corticosterone levels in mice with noise-induced depression.¹²⁰ tPBM also promoted neuroprotection effects from stress,^{122,123} reserpine,¹²⁴ underwater trauma,¹²⁵ and early AD-associated¹²⁶ depression in rat models. Specifically, Tanaka et al. found that tPBM (bright light) significantly promoted neurogenesis in the hippocampal CA1 region.²³⁹ Li et al. found that tPBM (810 nm) elevated ATP levels in the hippocampus in rats with a posttraumatic stress disorders model and modulated activated neurons expressing immediate-early genes, such as Arc and c-fos in the hippocampus and amygdala.¹²⁵ Mohammed et al. found that low-dose (80 mW) tPBM significantly improved depression-like behavior in reserpine-induced depression in rats. Using electro-corticography spectral analysis, the beneficial effects of tPBM (80 mW) also included the inhibition of abnormal elevation of the Delta frequency band and decline of the Beta-1 and Beta-2 bands.¹²⁴ Salehpour et al. observed that a 10-Hz pulsed NIR laser (808 nm) was as effective as the Citalopram treatment and was more effective than a red laser (660 nm) in improving depressive-like behaviors in rats. This was likely due to a better penetration depth and higher absorption by CCO of the 808 nm laser.¹²³

In clinical trials, Schiffer et al. found that tPBM (810 nm) significantly improved the Hamilton depression rating scale and Hamilton anxiety rating scale scores in depression patients. There was increased mean rCBF in both hemispheres.⁴² Cassano et al. confirmed the safety and effectiveness of tPBM (823 nm) in major depressive disorder patients.⁴³ In addition, Disner et al. found that applying tPBM to the right prefrontal lobe enhanced the therapeutic effects of attention bias modification.⁴¹ Kerppers et al. found that tPBM (945 nm) was also effective for the treatment of depression.⁴⁴ In conclusion, the above compelling evidence shows that tPBM could be a promising alternative method for the therapy of depression in routine clinical practice.

3.10 tPBM for Aging

Aging is a multifactorial biological process, manifested by a progressive functional decline at the molecular, cellular, tissue, and organ levels. In addition, aging often increases the susceptibility to neurodegenerative and cardiovascular diseases, diabetes, and even cancer.²⁴⁰ Epidemiological studies show that 11% of the world population was over 60 years old in 2016, and it was estimated that this proportion will increase to 22% by 2050.²⁴¹ Although people welcome the prospect of longer life, they hope that old age is healthy rather than accompanied by disease. Therefore, it is important to explore the physiological and pathological processes of aging and find effective anti-aging therapeutics in the increasingly aging society. Oxidative stress and mitochondrial dysfunction are two important factors associated with aging.²⁴² Specifically, the oxidative stress theory indicates that the imbalance between the production of pro-oxidants and the defense of anti-oxidants during aging leads to excessive production and accumulation of ROS, damages macromolecules (lipids, DNA, and proteins), and then induces cell aging and death.²⁴³ In addition, mitochondria are considered to be the main target of oxidative damage, the dysfunction of which during aging is associated with cognitive impairment.²⁴⁴ Furthermore, oxidative stress within mitochondria can lead to a vicious cycle in which damaged mitochondria produce increased amounts of ROS, leading in turn to progressive augmentation in damage. Therefore, improving mitochondria function and inhibiting oxidative stress may have beneficial effects on the pathology associated with aging (see Table S10 in the Supplementary Material).

In pre-clinical studies, Sadigh–Eteghad's team found that transcranial red (660 nm) and NIR (810 nm) light treatment significantly alleviated memory and cognitive decline, mitochondrial dysfunction, oxidative stress, and cell apoptosis in the D-galactose-induced aging mouse model.^{127–129} Importantly, they found that a medium dose (8 J/cm² per day) was more effective than a low dose (4 J/cm² per day) or a high dose (32 J/cm² per day). There was no significant difference between red light (660 nm) and NIR light (810 nm). They also observed that red light (660 nm) alleviated cognitive impairment and mitochondrial dysfunction in naturally aging mice (18 months old).¹³⁰ Moreover, the cognitive improvement effect was also observed by 1072 nm laser treatment in Ennaceur's team.¹³¹ Massri et al. found that tPBM (670 nm) had beneficial effects on the inhibition of age-related glial cell (e.g., astrocyte and microglia) proliferation.¹³² Cardoso's team found that aging primarily leads to a decrease of regional brain CCO activity and impairment of systems-level FC and that tPBM (810 nm) reversed these age-related effects.¹³³ Furthermore, they also observed that tPBM mitigated the inflammatory response and altered the intracellular signaling pathways linked to vascular function, cell survival, and glucose metabolism in the aged brain.^{134–136}

In clinical trials, Salgado et al. found in elderly women that, using transcranial Doppler ultrasound measurements, tPBM (627 nm) significantly increased the systolic and diastolic velocities of the left middle cerebral artery and the basilar artery and decreased the fluctuation index and resistance index.⁴⁶ In addition, Chan et al. demonstrated improved depressive state and cognitive function of the elderly after applying combined red (633 nm) and NIR (870 nm) LEDs.^{48,49} Moreover, Saucedo et al. found that the beneficial effects of tPBM (1064 nm) for the elderly were not limited to improving cognitive function but also included increasing resting-stateEEG alpha, beta, gamma power, and prefrontal blood-oxygen-level-dependent -fMRI activity.⁴⁵ They also found that the CCO level was elevated during the irradiation period, followed by a significant post-stimulation increase in oxygenated hemoglobin and a decrease in deoxygenated hemoglobin.⁴⁷ Qu et al. observed that 7-day repeated tPBM efficiently improved the working memory of healthy older adults, with the beneficial effects lasting at least 3 weeks.⁵⁰ The above results indicated that tPBM could be a promising candidate for age-related cognitive improvement. In the future, there will be an urgent need to explore the functional changes related to phototherapy on the aging brain, including molecular and electrophysiological measurements and optical imaging techniques.

3.11 tPBM for Epilepsy

Epilepsy is a lifelong condition characterized by spontaneous and recurrent seizures. Epilepsy affects 70 million people worldwide and entails a major burden in seizure-related disability, mortality, comorbidities, stigma, and costs.²³⁶ Currently, anti-seizure medicines are the prevailing treatment modality for most people with epilepsy and are usually accompanied by low response rates (~70%) and high recurrence rates (>50%).¹³⁷ Although invasive neuronal stimulation treatments, such as TMS and tDCS, have been established as palliative treatments for patients with drug-resistant epilepsy, the low effectiveness and the high adverse event rate limit their clinical application.¹³⁷ Therefore, an effective and non-invasive therapeutic for epilepsy is urgently needed. Recently, tPBM with NIR light has been proposed as a new alternative treatment for epilepsy in animal model experiments (see Table S11 in the Supplementary Material).

Radwan et al. observed that tPBM (830 nm) reverses pilocarpine induced neurochemical changes of amino acid neurotransmitters (e.g., increase of glutamate), and they found that such benefits could be due to the regulation of transaminase activity by laser irradiation.¹³⁸ Vogel et al. observed that tPBM (780 nm) reduces epileptiform discharges in post-stroke epilepsy rats. The authors reasoned that the beneficial effects of tPBM might be responsible for the disruption and mitigation of self-regulatory mechanisms and complex network reorganization.¹³⁹ Tsai et al. found that applying tPBM before inducing epilepsy attenuated pentylenetetrazole (PTZ)-induced severe seizures, status epileptics, and mortality in peripubertal rats. This could be explained by protecting hippocampal parvalbumin-positive interneurons from apoptosis and preserving the integrity of the parvalbumin-positive perisomatic inhibitory network.¹³⁷ They also found that tPBM (808 nm) mitigated neuroinflammation and glial cell activation in the hippocampus in a CCO-dependent manner in the PTZ-induced epilepsy rat model.¹⁴⁰ Hong et al. observed that tPBM could prevent seizure-induced neuronal degeneration.¹⁴¹ In summary, the above studies

demonstrated that tPBM with NIR light could be potential non-invasive and effective therapeutics for inhibiting epileptogenesis and alleviating epilepsy damage.

4 Conclusion

In this review, we discussed new discoveries of therapeutic effects of tPBM to shed light on tPBM-mediated mechanisms of treatment for brain diseases. We also highlighted emerging trends in tPBM application for modulation of the immune system of the brain and stimulation of the MLVs, thus opening a new niche in tPBM-immunotherapy of brain diseases and in the removal of waste and toxins from the CNS. We summarized the animal experiments and clinical trials in different brain diseases. This is expected to provide guidance for future experimental designs and clinical applications.

Among the main disadvantages of tPBM is the limited information about the optimal tPBM protocols for the effective therapy of specific brain diseases. Despite the large number of experimental and clinical studies, it remains unclear whether the tPBM effects on the energy, metabolic, hemodynamic, drainage, and immune processes of the brain are nonspecific or whether they depend on specific characteristics of brain diseases. A better understanding of the dosedependent effects of tPBM on the brain physiology and mechanisms of tPBM therapy will help to develop the optimal and personalized tPBM standards and guidelines of this therapy for brain diseases.

The light delivery strategy of tPBM is also important for the treatment of brain diseases. The majority of the light delivery approaches are transcranial or intranasal. However, photons are scattered by the scalp, skull, meninges, and cerebrospinal fluid. Thus, only a small number of photons can effectively the reach brain tissues. In fact, in most brain diseases, deeper areas of the CNS are also involved. For example, in patients with PD, the midbrain is characterized by the progressive, selective loss of dopaminergic neurons in the SNc region. Therefore, the development of innovative strategies to deliver light deeper, including, for example, modern lasers (e.g., 1267 nm) with deep penetration into brain tissues, will open a new era in tPBM for treatment of brain diseases.

Given that the BBB prevents drug delivery to the brain and significantly limits the advancement in the development of new pharmacological therapies for brain diseases, tPBM can be an important therapeutic approach for preventing or delaying neurological pathologies. It is important to note that some companies have ceased initiatives to finance the development of pharmacological treatments for AD. Therefore, tPBM-mediated stimulation of the MLVs and removal of toxins, including A β from the brain, might be an important method for the treatment of this disease. tPBM-stimulating effects on the mechanisms of lymphatic drainage and the brain immune system open pioneering strategy for neuroprotection of the CNS in various brain diseases.

The special advantages of tPBM include its safety, ease of use in clinical settings and at home, and cost effectiveness. This makes tPBM attractive in the market of medical devices. Therefore, a better understanding of the mechanisms of tPBM, the development of generally accepted guidelines for tPBM use in clinical practice, and the study of dose-dependent and personal tPBM needs/effects hold great promise for progress in the treatment of brain diseases.

Disclosures

The authors declare that there are no conflicts of interest related to this article.

Code and Data Availability

Data sharing is not applicable to this article, as no new data were created or analyzed.

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References

- Y. H. Wang et al., "Depression and anxiety in relation to cancer incidence and mortality: a systematic review and meta-analysis of cohort studies," *Mol. Psychiatry* 25(7), 1487–1499 (2020).
- C. M. Stinear et al., "Advances and challenges in stroke rehabilitation," *Lancet Neurol.* 19(4), 348–360 (2020).
- L. L. Barnes, "Alzheimer disease in African American individuals: increased incidence or not enough data?" Nat. Rev. Neurol. 18(1), 56–62 (2022).
- E. Bullmore and O. Sporns, "The economy of brain network organization," *Nat. Rev. Neurosci.* 13(5), 336–349 (2012).
- E. C. M. de Lange, "The mastermind approach to CNS drug therapy: translational prediction of human brain distribution, target site kinetics, and therapeutic effects," *Fluids Barriers CNS* 10(1), 12 (2013).
- 6. X. Dong, "Current strategies for brain drug delivery," Theranostics 8(6), 1481-1493 (2018).
- O. Lindvall and Z. Kokaia, "Stem cell therapy for human brain disorders," *Kidney Int.* 68(5), 1937–1939 (2005).
- M. J. Begemann et al., "Efficacy of non-invasive brain stimulation on cognitive functioning in brain disorders: a meta-analysis," *Psychol. Med.* 50(15), 2465–2486 (2020).
- M. Hasselblatt, H. Ehrenreich, and A. L. Sirén, "The brain erythropoietin system and its potential for therapeutic exploitation in brain disease," *J. Neurosurg. Anesthesiol.* 18(2), 132–138 (2006).
- W. Peng et al., "High-frequency terahertz waves disrupt Alzheimer's β-amyloid fibril formation," *eLight* 3, 18 (2023).
- C. C. Hernandez et al., "Neuroprotective effects of erythropoietin on neurodegenerative and ischemic brain diseases: the role of erythropoietin receptor," *Neural Regener. Res.* 12(9), 1381–1389 (2017).
- 12. M. M. Rahman et al., "Stem cell transplantation therapy and neurological disorders: current status and future perspectives," *Biology* **11**(1), 147 (2022).
- A. Singh et al., "Transcranial magnetic and direct current stimulation in the treatment of depression: basic mechanisms and challenges of two commonly used brain stimulation methods in interventional psychiatry," *Neuropsychobiology* **79**(6), 397–407 (2020).
- H. W. Chase et al., "Transcranial direct current stimulation: a roadmap for research, from mechanism of action to clinical implementation," *Mol. Psychiatry* 25(2), 397–407 (2020).
- V. Heiskanen and M. R. Hamblin, "Photobiomodulation: lasers vs. light emitting diodes?" *Photochem. Photobiol. Sci.* 17(8), 1003–1017 (2018).
- S. Farivar, T. Malekshahabi, and R. Shiari, "Biological effects of low level laser therapy," *J. Lasers Med. Sci.* 5(2), 58–62 (2014).
- F. Salehpour et al., "Photobiomodulation therapy and the glymphatic system: promising applications for augmenting the brain lymphatic drainage system," *Int. J. Mol. Sci.* 23(6), 2975 (2022).
- J. C. Rojas and F. Gonzalez-Lima, "Low-level light therapy of the eye and brain," *Eye Brain* 3, 49–67 (2011).
- F. Salehpour and M. R. Hamblin, "Photobiomodulation for Parkinson's disease in animal models: a systematic review," *Biomolecules* 10(4), 610 (2020).
- Y. Lampl et al., "Infrared laser therapy for ischemic stroke: a new treatment strategy: results of the NeuroThera Effectiveness and Safety Trial-1 (NEST-1)," *Stroke* 38(6), 1843–1849 (2007).
- J. A. Zivin et al., "Effectiveness and safety of transcranial laser therapy for acute ischemic stroke," *Stroke* 40(4), 1359–1364 (2009).
- M. A. Naeser et al., "Increased functional connectivity within intrinsic neural networks in chronic stroke following treatment with red/near-infrared transcranial photobiomodulation: case series with improved naming in aphasia," *Photobiomodulation Photomed. Laser Surg.* 38(2), 115–131 (2020).
- K. Estrada-Rojas and N.P. Cedeño Ortiz, "Increased improvement in speech-language skills after transcranial photobiomodulation plus speech-language therapy, compared to speech-language therapy alone: case report with aphasia," *Photobiomodulation Photomed. Laser Surg.* 41(5), 234–240 (2023).
- A. B. Stemer, B. N. Huisa, and J. A. Zivin, "The evolution of transcranial laser therapy for acute ischemic stroke, including a pooled analysis of NEST-1 and NEST-2," *Curr. Cardiol. Rep.* 12(1), 29–33 (2010).
- A. E. Saltmarche et al., "Significant improvement in cognition in mild to moderately severe dementia cases treated with transcranial plus intranasal photobiomodulation: case series report," *Photomed. Laser Surg.* 35(8), 432–441 (2017).
- L. L. Chao, "Effects of home photobiomodulation treatments on cognitive and behavioral function, cerebral perfusion, and resting-state functional connectivity in patients with dementia: a pilot trial," *Photobiomodulation Photomed. Laser Surg.* 37(3), 133–141 (2019).

- F. Salehpour, M. R. Hamblin, and J. O. DiDuro, "Rapid reversal of cognitive decline, olfactory dysfunction, and quality of life using multi-modality photobiomodulation therapy: case report," *Photobiomodulation Photomed. Laser Surg.* 37(3), 159–167 (2019).
- M. H. Berman et al., "Photobiomodulation with near infrared light helmet in a pilot, placebo controlled clinical trial in dementia patients testing memory and cognition," J. Neurol. Neurosci. 8(1), 176 (2017).
- 29. A. Liebert et al., "Improvements in clinical signs of Parkinson's disease using photobiomodulation: a prospective proof-of-concept study," *BMC Neurol.* **21**(1), 256 (2021).
- A. Liebert et al., "Remote photobiomodulation treatment for the clinical signs of Parkinson's disease: a case series conducted during COVID-19," *Photobiomodulation Photomed. Laser Surg.* 40(2), 112–122 (2022).
- C. L. Hamilton et al., ""Buckets": early observations on the use of red and infrared light helmets in Parkinson's disease patients," *Photobiomodulation Photomed. Laser Surg.* 37(10), 615–622 (2019).
- C. T. Hong et al., "Effects of concomitant use of hydrogen water and photobiomodulation on Parkinson disease: a pilot study," *Medicine* 100(4), e24191 (2021).
- C. McGee et al., "A randomized placebo-controlled study of a transcranial photobiomodulation helmet in Parkinson's disease: post-hoc analysis of motor outcomes," J. Clin. Med. 12(8), 2846 (2023).
- 34. A. Liebert et al., "Improvements in the clinical signs of Parkinson's disease using photobiomodulation: a 3-year follow-up case series," *Med. Res. Arch.* **11**(3), 256 (2023).
- 35. L. L. Chao et al., "Changes in brain function and structure after self-administered home photobiomodulation treatment in a concussion case," *Front. Neurol.* **11**, 952 (2020).
- M. A. Naeser et al., "Significant improvements in cognitive performance post-transcranial, red/nearinfrared light-emitting diode treatments in chronic, mild traumatic brain injury: open-protocol study," *J. Neurotrauma* 31(11), 1008–1017 (2014).
- 37. M. A. Naeser et al., "Transcranial, red/near-infrared light-emitting diode therapy to improve cognition in chronic traumatic brain injury," *Photomed. Laser Surg.* **34**(12), 610–626 (2016).
- M. A. Naeser et al., "Improved cognitive function after transcranial, light-emitting diode treatments in chronic, traumatic brain injury: two case reports," *Photomed. Laser Surg.* 29(5), 351–358 (2011).
- M. G. Figueiro Longo et al., "Effect of transcranial low-level light therapy vs sham therapy among patients with moderate traumatic brain injury: a randomized clinical trial," *JAMA Netw. Open* 3(9), e2017337 (2020).
- H. Nawashiro et al., "Focal increase in cerebral blood flow after treatment with near-infrared light to the forehead in a patient in a persistent vegetative state," *Photomed. Laser Surg.* 30(4), 231–233 (2012).
- S. G. Disner, C. G. Beevers, and F. Gonzalez-Lima, "Transcranial laser stimulation as neuroenhancement for attention bias modification in adults with elevated depression symptoms," *Brain Stimul.* 9(5), 780–787 (2016).
- 42. F. Schiffer et al., "Psychological benefits 2 and 4 weeks after a single treatment with near infrared light to the forehead: a pilot study of 10 patients with major depression and anxiety," *Behav. Brain Funct.* 5, 46 (2009).
- P. Cassano et al., "Transcranial photobiomodulation for the treatment of major depressive disorder. The ELATED-2 pilot trial," *Photomed. Laser Surg.* 36(12), 634–646 (2018).
- F. K. Kerppers et al., "Study of transcranial photobiomodulation at 945-nm wavelength: anxiety and depression," *Lasers Med. Sci.* 35(9), 1945–1954 (2020).
- E. Vargas et al., "Beneficial neurocognitive effects of transcranial laser in older adults," *Lasers Med. Sci.* 32(5), 1153–1162 (2017).
- A. S. Salgado et al., "The effects of transcranial LED therapy (TCLT) on cerebral blood flow in the elderly women," *Lasers Med. Sci.* 30(1), 339–346 (2015).
- C. L. Saucedo et al., "Transcranial laser stimulation: mitochondrial and cerebrovascular effects in younger and older healthy adults," *Brain Stimul.* 14(2), 440–449 (2021).
- A. S. Chan et al., "Photobiomodulation improves the frontal cognitive function of older adults," *Int. J. Geriatr. Psychiatry* 34(2), 369–377 (2019).
- 49. A. S. Chan et al., "Photobiomodulation enhances memory processing in older adults with mild cognitive impairment: a functional near-infrared spectroscopy study," J. Alzheimer's Dis. 83, 1471–1480 (2021).
- X. Qu et al., "Repeated transcranial photobiomodulation improves working memory of healthy older adults: behavioral outcomes of poststimulation including a three-week follow-up," *Neurophotonics* 9(3), 035005 (2022).
- H. I. Lee et al., "Pre-conditioning with transcranial low-level light therapy reduces neuroinflammation and protects blood-brain barrier after focal cerebral ischemia in mice," *Restorative Neurol. Neurosci.* 34(2), 201–214 (2016).
- 52. H. Kim et al., "Benefits of a skull-interfaced flexible and implantable multilight emitting diode array for photobiomodulation in ischemic stroke," *Adv. Sci.* **9**(11), e2104629 (2022).
- H. I. Lee et al., "Pretreatment with light-emitting diode therapy reduces ischemic brain injury in mice through endothelial nitric oxide synthase-dependent mechanisms," *Biophys. Res. Commun.* 486(4), 945–950 (2017).

- 54. H. I. Lee et al., "Low-level light emitting diode (LED) therapy suppresses inflammasome-mediated brain damage in experimental ischemic stroke," *J. Biophotonics* **10**(11), 1502–1513 (2017).
- 55. H. I. Lee et al., "Low-level light emitting diode therapy promotes long-term functional recovery after experimental stroke in mice," *J. Biophotonics* **10**(12), 1761–1771 (2017).
- M. C. Leung et al., "Treatment of experimentally induced transient cerebral ischemia with low energy laser inhibits nitric oxide synthase activity and up-regulates the expression of transforming growth factor-beta 1," *Lasers Surg. Med.* 31(4), 283–288 (2002).
- D. D. S. Vogel et al., "Transcranial low-level laser therapy in an *in vivo* model of stroke: relevance to the brain infarct, microglia activation and neuroinflammation," *J. Biophotonics* 14(6), e202000500 (2021).
- A. Oron et al., "Low-level laser therapy applied transcranially to rats after induction of stroke significantly reduces long-term neurological deficits," *Stroke* 37(10), 2620–2624 (2006).
- R. Wang et al., "Photobiomodulation for global cerebral ischemia: targeting mitochondrial dynamics and functions," *Mol. Neurobiol.* 56(3), 1852–1869 (2019).
- 60. L. Yang et al., "Photobiomodulation therapy promotes neurogenesis by improving post-stroke local microenvironment and stimulating neuroprogenitor cells," *Exp. Neurol.* **299**, 86–96 (2018).
- E. G. de Jesus Fonseca et al., "Study of transcranial therapy 904 nm in experimental model of stroke," *Lasers Med. Sci.* 34(8), 1619–1625 (2019).
- L. DeTaboada et al., "Transcranial application of low-energy laser irradiation improves neurological deficits in rats following acute stroke," *Lasers Surg. Med.* 38(1), 70–73 (2006).
- P. A. Lapchak, J. Wei, and J. A. Zivin, "Transcranial infrared laser therapy improves clinical rating scores after embolic strokes in rabbits," *Stroke* 35(8), 1985–1988 (2004).
- 64. B. N. Huisa et al., "Incremental treatments with laser therapy augments good behavioral outcome in the rabbit small clot embolic stroke model," *Lasers Med. Sci.* 28(4), 1085–1089 (2013).
- P. A. Lapchak and L. De Taboada, "Transcranial near infrared laser treatment (NILT) increases cortical adenosine-5'-triphosphate (ATP) content following embolic strokes in rabbits," *Brain Res.* 1306, 100–105 (2010).
- D. M. Meyer, Y. Chen, and J. A. Zivin, "Dose-finding study of phototherapy on stroke outcome in a rabbit model of ischemic stroke," *Neurosci. Lett.* 630, 254–258 (2016).
- P. A. Lapchak et al., "Transcranial near-infrared light therapy improves motor function following embolic strokes in rabbits: an extended therapeutic window study using continuous and pulse frequency delivery modes," *Neuroscience* 148(4), 907–914 (2007).
- 68. P. A. Lapchak et al., "Safety profile of transcranial near-infrared laser therapy administered in combination with thrombolytic therapy to embolized rabbits," *Stroke* 39(11), 3073–3078 (2008).
- 69. P. A. Lapchak and P. D. Boitano, "A novel method to promote behavioral improvement and enhance mitochondrial function following an embolic stroke," *Brain Res.* **1646**, 125–131 (2016).
- W. Hacke et al., "Transcranial laser therapy in acute stroke treatment: results of neurothera effectiveness and safety trial 3, a phase III clinical end point device trial," *Stroke* 45(11), 3187–3193 (2014).
- L. Yang et al., "Effects of prenatal photobiomodulation treatment on neonatal hypoxic ischemia in rat offspring," *Theranostics* 11(3), 1269–1294 (2021).
- L. D. Tucker et al., "Photobiomodulation therapy attenuates hypoxic-ischemic injury in a neonatal rat model," J. Mol. Neurosci. 65(4), 514–526 (2018).
- 73. L. Yang et al., "Photobiomodulation preconditioning prevents cognitive impairment in a neonatal rat model of hypoxia-ischemia," *J. Biophotonics* **12**(6), e201800359 (2019).
- D. Li et al., "Photostimulation of brain lymphatics in male newborn and adult rodents for therapy of intraventricular hemorrhage," *Nat. Commun.* 14(1), 6104 (2023).
- L. Tao et al., "Microglia modulation with 1070-nm light attenuates Abeta burden and cognitive impairment in Alzheimer's disease mouse model," *Light Sci. Appl.* 10(1), 179 (2021).
- Y. Lu et al., "Low-level laser therapy for beta amyloid toxicity in rat hippocampus," *Neurobiol. Aging* 49, 165–182 (2017).
- L. Yang et al., "Non-invasive photobiomodulation treatment in an Alzheimer disease-like transgenic rat model," *Theranostics* 12(5), 2205–2231 (2022).
- O. Semyachkina-Glushkovskaya et al., "Night photostimulation of clearance of beta-amyloid from mouse brain: new strategies in preventing Alzheimer's disease," *Cells* 10(12), 3289 (2021).
- E. Zinchenko et al., "Pilot study of transcranial photobiomodulation of lymphatic clearance of beta-amyloid from the mouse brain: breakthrough strategies for non-pharmacologic therapy of Alzheimer's disease," *Biomed. Opt. Express* 10(8), 4003–4017 (2019).
- Z. Zhang et al., "Activation of PKA/SIRT1 signaling pathway by photobiomodulation therapy reduces Abeta levels in Alzheimer's disease models," *Aging Cell* 19(1), e13054 (2020).
- X. Yue et al., "New insight into Alzheimer's disease: light reverses Aβ-obstructed interstitial fluid flow and ameliorates memory decline in APP/PS1 mice," *Alzheimer's Dementia Transl. Res. Clin. Interv.* 5, 671–684 (2019).

- Q. Shen et al., "Photobiomodulation suppresses JNK3 by activation of ERK/MKP7 to attenuate AMPA receptor endocytosis in Alzheimer's disease," *Aging Cell* 20(1), e13289 (2021).
- S. L. Grillo et al., "Non-invasive infra-red therapy (1072 nm) reduces β-amyloid protein levels in the brain of an Alzheimer's disease mouse model, TASTPM," J. Photochem. Photobiol. B Biol. 123, 13–22 (2013).
- V. E. Shaw et al., "Neuroprotection of midbrain dopaminergic cells in MPTP-treated mice after near-infrared light treatment," J. Comp. Neurol. 518(1), 25–40 (2010).
- C. Peoples et al., "Photobiomodulation enhances nigral dopaminergic cell survival in a chronic MPTP mouse model of Parkinson's disease," *Parkinsonism Relat. Disorder* 18(5), 469–476 (2012).
- C. Peoples et al., "Survival of dopaminergic amacrine cells after near-infrared light treatment in MPTPtreated mice," *ISRN Neurol.* 2012, 850150 (2012).
- V. E. Shaw et al., "Patterns of cell activity in the subthalamic region associated with the neuroprotective action of near-infrared light treatment in MPTP-treated mice," *Parkinsons Dis.* 2012, 296875 (2012).
- F. Reinhart et al., "810 nm near-infrared light offers neuroprotection and improves locomotor activity in MPTP-treated mice," *Neurosci. Res.* 92, 86–90 (2015).
- C. Moro et al., "Photobiomodulation preserves behaviour and midbrain dopaminergic cells from MPTP toxicity: evidence from two mouse strains," *BMC Neurosci.* 14(1), 40 (2013).
- M. San Miguel et al., "Photobiomodulation mitigates cerebrovascular leakage induced by the Parkinsonian neurotoxin MPTP," *Biomolecules* 9(10), 564 (2019).
- J. A. O'Brien and P. J. Austin, "Effect of photobiomodulation in rescuing lipopolysaccharide-induced dopaminergic cell loss in the male Sprague-Dawley rat," *Biomolecules* 9(8), 381 (2019).
- 92. A. S. Salgado et al., "Effects of light emitting diode and low-intensity light on the immunological process in a model of Parkinson's disease," *Med. Res. Arch.* **4**(8) (2016).
- A. Oueslati et al., "Photobiomodulation suppresses alpha-synuclein-induced toxicity in an AAV-based rat genetic model of Parkinson's disease," *PLoS One* 10(10), e0140880 (2015).
- S. Purushothuman et al., "The impact of near-infrared light on dopaminergic cell survival in a transgenic mouse model of parkinsonism," *Brain Res.* 1535, 61–70 (2013).
- F. Darlot et al., "Near-infrared light is neuroprotective in a monkey model of Parkinson disease," Ann. Neurol. 79(1), 59–75 (2016).
- D. M. Johnstone et al., "Indirect application of near infrared light induces neuroprotection in a mouse model of Parkinsonism: an abscopal neuroprotective effect," *Neuroscience* 274, 93–101 (2014).
- T. Silva et al., "Effects of photobiomodulation on interleukin-10 and nitrites in individuals with relapsingremitting multiple sclerosis: randomized clinical trial," *PLoS One* 15(4), e0230551 (2020).
- K. C. N. Duarte et al., "Low-level laser therapy modulates demyelination in mice," J. Photochem. Photobiol. B Biol. 189, 55–65 (2018).
- T. Silva et al., "Effect of photobiomodulation on fatigue in individuals with relapsing-remitting multiple sclerosis: a pilot study," *Lasers Med. Sci.* 37(8), 3107–3113 (2022).
- R. O. Esenaliev et al., "Nano-pulsed laser therapy is neuroprotective in a rat model of blast-induced neurotrauma," J. Neurotrauma 35(13), 1510–1522 (2018).
- 101. T. Ando et al., "Comparison of therapeutic effects between pulsed and continuous wave 810-nm wavelength laser irradiation for traumatic brain injury in mice," *PLoS One* **6**(10), e26212 (2011).
- Q. Wu et al., "Low-level laser therapy for closed-head traumatic brain injury in mice: effect of different wavelengths," *Lasers Surg. Med.* 44(3), 218–226 (2012).
- W. Xuan et al., "Transcranial low-level laser therapy improves neurological performance in traumatic brain injury in mice: effect of treatment repetition regimen," *PLoS One* 8(1), e53454 (2013).
- W. Xuan et al., "Transcranial low-level laser therapy enhances learning, memory, and neuroprogenitor cells after traumatic brain injury in mice," J. Biomed. Opt. 19(10), 108003 (2014).
- W. Xuan et al., "Low-level laser therapy for traumatic brain injury in mice increases brain derived neurotrophic factor (BDNF) and synaptogenesis," J. Biophotonics 8(6), 502–511 (2015).
- W. Xuan, L. Huang, and M. R. Hamblin, "Repeated transcranial low-level laser therapy for traumatic brain injury in mice: biphasic dose response and long-term treatment outcome," *J. Biophotonics* 9(11–12), 1263–1272 (2016).
- D. Shemesh et al., "Quantification of the effectivity of laser therapy shortly following brain injury via dualwavelength laser imaging," *Opt. Laser Technol.* 145, 107506 (2022).
- 108. Q. Zhang et al., "Low-level laser therapy effectively prevents secondary brain injury induced by immediate early responsive gene X-1 deficiency," *J. Cereb. Blood Flow Metab.* **34**(8), 1391–1401 (2014).
- T. Dong et al., "Low-level light in combination with metabolic modulators for effective therapy of injured brain," J. Cereb. Blood Flow Metab. 35(9), 1435–1444 (2015).
- E. Mocciaro et al., "Non-invasive transcranial nano-pulsed laser therapy ameliorates cognitive function and prevents aberrant migration of neural progenitor cells in the hippocampus of rats subjected to traumatic brain injury," *J. Neurotrauma* 37(8), 1108–1123 (2020).

- 111. B. J. Quirk et al., "Near-infrared photobiomodulation in an animal model of traumatic brain injury: improvements at the behavioral and biochemical levels," *Photomed. Laser Surg.* **30**(9), 523–529 (2012).
- 112. J. Khuman et al., "Low-level laser light therapy improves cognitive deficits and inhibits microglial activation after controlled cortical impact in mice," J. Neurotrauma 29(2), 408–417 (2012).
- M. S. Moreira et al., "Effect of phototherapy with low intensity laser on local and systemic immunomodulation following focal brain damage in rat," J. Photochem. Photobiol. B Biol. 97(3), 145–151 (2009).
- A. Oron et al., "Low-level laser therapy applied transcranially to mice following traumatic brain injury significantly reduces long-term neurological deficits," *J. Neurotrauma* 24(4), 651–656 (2007).
- 115. A. Oron et al., "Near infrared transcranial laser therapy applied at various modes to mice following traumatic brain injury significantly reduces long-term neurological deficits," *J. Neurotrauma* 29(2), 401–407 (2012).
- B. Yang et al., "Photobiomodulation therapy for repeated closed head injury in rats," *J. Biophotonics* 13(2), e201960117 (2020).
- 117. M. A. Naeser et al., "Transcranial photobiomodulation treatment: significant improvements in four ex-football players with possible chronic traumatic encephalopathy," *J. Alzheimer's Dis. Rep.* 7, 77–105 (2023).
- F. Salehpour et al., "Transcranial near-infrared photobiomodulation attenuates memory impairment and hippocampal oxidative stress in sleep-deprived mice," *Brain Res.* 1682, 36–43 (2018).
- F. Salehpour et al., "Near-infrared photobiomodulation combined with coenzyme Q10 for depression in a mouse model of restraint stress: reduction in oxidative stress, neuroinflammation, and apoptosis," *Brain Res. Bull.* 144, 213–222 (2019).
- 120. N. Farazi et al., "Synergistic effects of combined therapy with transcranial photobiomodulation and enriched environment on depressive- and anxiety-like behaviors in a mice model of noise stress," *Lasers Med. Sci.* 37(2), 1181–1191 (2022).
- Z. Xu et al., "Low-level laser irradiation improves depression-like behaviors in mice," *Mol. Neurobiol.* 54(6), 4551–4559 (2017).
- 122. X. Wu et al., "Pulsed light irradiation improves behavioral outcome in a rat model of chronic mild stress," *Lasers Surg. Med.* **44**(3), 227–232 (2012).
- 123. F. Salehpour et al., "Therapeutic effects of 10-Hz pulsed wave lasers in rat depression model: a comparison between near-infrared and red wavelengths," *Lasers Surg. Med.* 48(7), 695–705 (2016).
- H. S. Mohammed, "Transcranial low-level infrared laser irradiation ameliorates depression induced by reserpine in rats," *Lasers Med. Sci.* 31(8), 1651–1656 (2016).
- Y. Li et al., "Transcranial photobiomodulation prevents PTSD-like comorbidities in rats experiencing underwater trauma," *Transl. Psychiatry* 11(1), 270 (2021).
- L. Yang et al., "Photobiomodulation therapy attenuates anxious-depressive-like behavior in the TgF344 rat model," J. Alzheimer's Dis. 83(4), 1415–1429 (2021).
- 127. F. Salehpour et al., "Transcranial low-level laser therapy improves brain mitochondrial function and cognitive impairment in D-galactose-induced aging mice," *Neurobiol. Aging* 58, 140–150 (2017).
- 128. F. Salehpour et al., "Photobiomodulation and coenzyme Q10 treatments attenuate cognitive impairment associated with model of transient global brain ischemia in artificially aged mice," *Front. Cell. Neurosci.* 13, 74 (2019).
- L. Hosseini et al., "Effect of transcranial near-infrared photobiomodulation on cognitive outcomes in D-galactose/AlCl3 induced brain aging in BALB/c mice," *Lasers Med. Sci.* 37(3), 1787–1798 (2022).
- 130. F. Salehpour et al., "A protocol for transcranial photobiomodulation therapy in mice," *J. Visual. Exp.* **141**, e59076 (2018).
- S. Michalikova et al., "Emotional responses and memory performance of middle-aged CD1 mice in a 3D maze: effects of low infrared light," *Neurobiol. Learn. Memory* 89(4), 480–488 (2008).
- N. El Massri et al., "Photobiomodulation reduces gliosis in the basal ganglia of aged mice," *Neurobiol. Aging* 66, 131–137 (2018).
- F. d. S. Cardoso et al., "Photobiomodulation of cytochrome c oxidase by chronic transcranial laser in young and aged brains," *Front. Neurosci.* 16, 818005 (2022).
- 134. F. d. S. Cardoso et al., "Transcranial laser photobiomodulation improves intracellular signaling linked to cell survival, memory and glucose metabolism in the aged brain: a preliminary study," *Front. Cell. Neurosci.* 15, 683127 (2021).
- 135. F. d. S. Cardoso et al., "Photobiomodulation improves the inflammatory response and intracellular signaling proteins linked to vascular function and cell survival in the brain of aged rats," *Mol. Neurobiol.* 59(1), 420–428 (2022).
- 136. F. d. S. Cardoso et al., "Improved spatial memory and neuroinflammatory profile changes in aged rats submitted to photobiomodulation therapy," *Cell. Mol. Neurobiol.* 42(6), 1875–1886 (2022).

- C. M. Tsai, S. F. Chang, and H. Chang, "Transcranial photobiomodulation attenuates pentylenetetrazoleinduced status epilepticus in peripubertal rats," *J. Biophotonics* 13(8), e202000095 (2020).
- N. M. Radwan et al., "Effect of infrared laser irradiation on amino acid neurotransmitters in an epileptic animal model induced by pilocarpine," *Photomed. Laser Surg.* 27(3), 401–409 (2009).
- D. D. S. Vogel et al., "Repetitive transcranial photobiomodulation but not long-term omega-3 intake reduces epileptiform discharges in rats with stroke-induced epilepsy," J. Biophotonics 14(1), e202000287 (2021).
- 140. C. M. Tsai et al., "Transcranial photobiomodulation (808 nm) attenuates pentylenetetrazole-induced seizures by suppressing hippocampal neuroinflammation, astrogliosis, and microgliosis in peripubertal rats," *Neurophotonics* 9(1), 015006 (2022).
- N. Hong et al., "Photobiomodulation regulates adult neurogenesis in the hippocampus in a status epilepticus animal model," *Sci. Rep.* 12(1), 15246 (2022).
- 142. T. Li et al., "Optimize illumination parameter of low-level laser therapy for hemorrhagic stroke by Monte Carlo simulation on visible human dataset," *IEEE Photonics J.* **10**(3), 6100409 (2018).
- 143. S. Kashiwagi et al., "Photobiomodulation and nitric oxide signaling," Nitric Oxide 130, 58-68 (2023).
- E. A. Genina et al., "Optical properties of brain tissues at the different stages of glioma development in rats: pilot study," *Biomed. Opt. Express* 10(10), 5182–5197 (2019).
- 145. D. Li et al., "Visible-near infrared-II skull optical clearing window for *in vivo* cortical vasculature imaging and targeted manipulation," *J. Biophotonics* **13**(10), e202000142 (2020).
- N. G. Horton et al., "In vivo three-photon microscopy of subcortical structures within an intact mouse brain," Nat. Photonics 7(3), 205–209 (2013).
- O. Semyachkina-Glushkovskaya et al., "Photobiomodulation of lymphatic drainage and clearance: perspective strategy for augmentation of meningeal lymphatic functions," *Biomed. Opt. Express* 11(2), 725–734 (2020).
- 148. A. Khokhlova et al., "The photobiomodulation of vital parameters of the cancer cell culture by low dose of near-IR laser irradiation," *IEEE J. Sel. Top. Quantum Electron.* 25(1), 7201510 (2019).
- 149. A. Khokhlova et al., "The light-oxygen effect in biological cells enhanced by highly localized surface plasmon-polaritons," *Sci. Rep.* **9**(1), 18435 (2019).
- C. P. Stanley et al., "Singlet molecular oxygen regulates vascular tone and blood pressure in inflammation," *Nature* 566(7745), 548–552 (2019).
- 151. G. E. Ronsein et al., "Tryptophan oxidation by singlet molecular oxygen [O2 (1Δg)]: mechanistic studies using 18O-labeled hydroperoxides, mass spectrometry, and light emission measurements," *Chem. Res. Toxicol.* 21(6), 1271–1283 (2008).
- Y. Wang et al., "Kynurenine is an endothelium-derived relaxing factor produced during inflammation," *Nat. Med.* 16(3), 279–285 (2010).
- 153. J. T. Hashmi et al., "Effect of pulsing in low-level light therapy," Lasers Surg. Med. 42(6), 450–466 (2010).
- P. Brondon, I. Stadler, and R. J. Lanzafame, "Pulsing influences photoradiation outcomes in cell culture," *Lasers Surg. Med.* 41(3), 222–226 (2009).
- 155. B. Shariati et al., "Method for tissue clearing: temporal tissue optical clearing," *Biomed. Opt. Express* **13**(8), 4222–4235 (2022).
- 156. D. Galiakhmetova et al., "Ultra-short laser pulses propagation through mouse head tissues: experimental and computational study," *IEEE J. Sel. Top. Quantum Electron.* **29**, 7200311 (2023).
- 157. S. V. Moskvin, "Only lasers can be used for low level laser therapy," *Biomedicine* 7(4), 22 (2017).
- K. Sato et al., "Comparative effectiveness of light emitting diodes (LEDs) and lasers in near infrared photoimmunotherapy," *Oncotarget* 7(12), 14324–14335 (2016).
- D. Litscher and G. Litscher, "Laser therapy and dementia: a database analysis and future aspects on LEDbased systems," *Int. J. Photoenergy* 2014, 268354 (2014).
- 160. P. Suchonwanit, N. Chalermroj, and S. Khunkhet, "Low-level laser therapy for the treatment of androgenetic alopecia in Thai men and women: a 24-week, randomized, double-blind, sham device-controlled trial," *Lasers Med. Sci.* 34(6), 1107–1114 (2019).
- R. A. Mussttaf, D. F. L. Jenkins, and A. N. Jha, "Assessing the impact of low level laser therapy (LLLT) on biological systems: a review," *Int. J. Radiat. Biol.* 95(2), 120–143 (2019).
- 162. M. R. Hamblin, "Shining light on the head: photobiomodulation for brain disorders," *BBA Clinical* 6, 113–124 (2016).
- 163. I. Golovynska et al., "Macrophages modulated by red/NIR light: phagocytosis, cytokines, mitochondrial activity, Ca²⁺ influx, membrane depolarization and viability," *Photochem. Photobiol.* 98(2), 484–497 (2022).
- 164. X. Wang et al., "Up-regulation of cerebral cytochrome-c-oxidase and hemodynamics by transcranial infrared laser stimulation: a broadband near-infrared spectroscopy study," J. Cereb. Blood Flow Metab. 37(12), 3789–3802 (2017).
- T. Pruitt et al., "Transcranial photobiomodulation (tPBM) with 1,064-nm laser to improve cerebral metabolism of the human brain *in vivo*," *Lasers Surg. Med.* 52(9), 807–813 (2020).

- 166. T. Pruitt et al., "Photobiomodulation at different wavelengths boosts mitochondrial redox metabolism and hemoglobin oxygenation: lasers vs. light-emitting diodes *in vivo*," *Metabolites* 12(2), 103 (2022).
- 167. R. H. Michael and N. D. Tatiana, "Mechanisms of low level light therapy," *Proc. SPIE* 6140, 614001 (2006).
- S. G. Sokolovski et al., "Singlet oxygen stimulates mitochondrial bioenergetics in brain cells," *Free Radical Biol. Med.* 163, 306–313 (2021).
- X. Wang et al., "Transcranial photobiomodulation with 1064-nm laser modulates brain electroencephalogram rhythms," *Neurophotonics* 6(2), 025013 (2019).
- 170. X. Wang et al., "Transcranial photobiomodulation and thermal stimulation induce distinct topographies of EEG alpha and beta power changes in healthy humans," *Sci. Rep.* 11(1), 18917 (2021).
- N. C. D. Truong et al., "Enhancement of frequency-specific hemodynamic power and functional connectivity by transcranial photobiomodulation in healthy humans," *Front. Neurosci.* 16, 896502 (2022).
- F. Tian et al., "Transcranial laser stimulation improves human cerebral oxygenation," *Lasers Surg. Med.* 48(4), 343–349 (2016).
- 173. Y. Zhao, P. M. Vanhoutte, and S. W. Leung, "Vascular nitric oxide: beyond eNOS," J. Pharmacol. Sci. 129(2), 83–94 (2015).
- 174. C. F. Rizzi et al., "Effects of low-level laser therapy (LLLT) on the nuclear factor (NF)-kappaB signaling pathway in traumatized muscle," *Lasers Surg. Med.* 38(7), 704–713 (2006).
- 175. T. Zhou, T. Y. Ohulchanskyy, and J. Qu, "Effect of NIR light on the permeability of the blood-brain barriers in *in vitro* models," *Biomed. Opt. Express* **12**(12), 7544–7555 (2021).
- 176. A. E. Mohamed et al., "Femtosecond laser attenuates oxidative stress, inflammation, and liver fibrosis in rats: possible role of PPARgamma and Nrf2/HO-1 signaling," *Life Sci.* 307, 120877 (2022).
- 177. Y. Y. Huang et al., "Low-level laser therapy (LLLT) reduces oxidative stress in primary cortical neurons in vitro," *J. Biophotonics* 6(10), 829–838 (2013).
- M. R. Hamblin, "Mechanisms and applications of the anti-inflammatory effects of photobiomodulation," *AIMS Biophys.* 4(3), 337–361 (2017).
- S. Da Mesquita, Z. Fu, and J. Kipnis, "The meningeal lymphatic system: a new player in neurophysiology," *Neuron* 100(2), 375–388 (2018).
- L. F. de Freitas and M. R. Hamblin, "Proposed mechanisms of photobiomodulation or low-level light therapy," *IEEE J. Sel. Top. Quantum Electron.* 22(3), 348–364 (2016).
- C. Moro et al., "The effect of photobiomodulation on the brain during wakefulness and sleep," *Front. Neurosci.* 16, 942536 (2022).
- O. Semyachkina-Glushkovskaya et al., "Photomodulation of lymphatic delivery of liposomes to the brain bypassing the blood-brain barrier: new perspectives for glioma therapy," *Nanophotonics* 10(12), 3215–3227 (2021).
- O. Semyachkina-Glushkovskaya et al., "Biophotonic strategies of measurement and stimulation of the cranial and the extracranial lymphatic drainage function," *IEEE J. Sel. Top. Quantum Electron.* 27(4), 7400313 (2021).
- O. Semyachkina-Glushkovskaya et al., "Photostimulation of cerebral and peripheral lymphatic functions," *Transl. Biophotonics* 2(1–2), e201900036 (2020).
- 185. E. Zinchenko et al., "Photostimulation of extravasation of beta-amyloid through the model of blood-brain barrier," *Electronics* **9**(6), 1056 (2020).
- T. I. Karu, L. V. Pyatibrat, and N. I. Afanasyeva, "Cellular effects of low power laser therapy can be mediated by nitric oxide," *Lasers Surg. Med.* 36(4), 307–314 (2005).
- F. Murad, "Discovery of some of the biological effects of nitric oxide and its role in cell signaling," *Biosci. Rep.* 24(4–5), 452–474 (2005).
- D. Weihrauch et al., "Red light stimulates vasodilation through extracellular vesicle trafficking," J. Photochem. Photobiol. B Biol. 220, 112212 (2021).
- J. P. Scallan et al., "Lymphatic pumping: mechanics, mechanisms and malfunction," J. Physiol. 594(20), 5749–5768 (2016).
- E. I. Galanzha, V. V. Tuchin, and V. P. Zharov, "Advances in small animal mesentery models for *in vivo* flow cytometry, dynamic microscopy, and drug screening," *World J. Gastroenterol.* 13(2), 192–218 (2007).
- G. Brill et al., "Influence of low-power laser irradiation on lymph microcirculation during increased NO production," *Proc. SPIE* 3726, 157–162 (1999).
- Y. Uozumi et al., "Targeted increase in cerebral blood flow by transcranial near-infrared laser irradiation," *Lasers Surg. Med.* 42(6), 566–576 (2010).
- E. L. Urquhart et al., "Transcranial photobiomodulation-induced changes in human brain functional connectivity and network metrics mapped by whole-head functional near-infrared spectroscopy *in vivo*," *Biomed. Opt. Express* 11(10), 5783–5799 (2020).
- 194. X. Wang et al., "Impact of heat on metabolic and hemodynamic changes in transcranial infrared laser stimulation measured by broadband near-infrared spectroscopy," *Neurophotonics* 5(1), 011004 (2017).

- 195. A. Shanshool and V. Tuchin, "Improved optical access of laser radiation to the brain using optical clearing and transparent cranial implants: review," *Proc. SPIE* **11845**, 118450W (2021).
- 196. N. Duggett and P. Chazot, "Low-intensity light therapy (1068 nm) protects CAD neuroblastoma cells from β -amyloid-mediated cell death," *Biol. Med.* **1**, 103 (2014).
- 197. C. Meng, Z. He, and D. Xing, "Low-level laser therapy rescues dendrite atrophy via upregulating BDNF expression: implications for Alzheimer's disease," J. Neurosci. 33(33), 13505–13517 (2013).
- 198. Y. C. Yun et al., "Laser acupuncture exerts neuroprotective effects via regulation of Creb, Bdnf, Bcl-2, and Bax gene expressions in the hippocampus," *Evid. Based Compl. Alternat. Med.* **2017**, 7181637 (2017).
- A. Telerman et al., "Induction of hippocampal neurogenesis by a tolerogenic peptide that ameliorates lupus manifestations," J. Neuroimmunol. 232(1–2), 151–157 (2011).
- S. G. Sokolovski et al., "Infrared laser pulse triggers increased singlet oxygen production in tumour cells," *Sci. Rep.* 3, 3484 (2013).
- S. D. Zakharov and A. V. Ivanov, "Light-oxygen effect in cells and its potential applications in tumour therapy (review)," *Quantum Electron.* 29(12), 1031 (1999).
- G. Minniti, G. Lombardi, and S. Paolini, "Glioblastoma in elderly patients: current management and future perspectives," *Cancers* 11(3), 336 (2019).
- X. Hu et al., "Meningeal lymphatic vessels regulate brain tumor drainage and immunity," *Cell Res.* 30(3), 229–243 (2020).
- 204. M. Kanamori and J. Kipnis, "Meningeal lymphatics "drain" brain tumors," *Cell Res.* **30**(3), 191–192 (2020).
- 205. Q. Ma et al., "Lymphatic outflow of cerebrospinal fluid is reduced in glioma," Sci. Rep. 9(1), 14815 (2019).
- E. S. Norton et al., "Glioblastoma disrupts the ependymal wall and extracellular matrix structures of the subventricular zone," *Fluids Barriers CNS* 19(1), 58 (2022).
- Y. L. Lan et al., "Update on the current knowledge of lymphatic drainage system and its emerging roles in glioma management," *Immunology* 168(2), 233–247 (2023).
- E. Song et al., "VEGF-C-driven lymphatic drainage enables immunosurveillance of brain tumours," *Nature* 577(7792), 689–694 (2020).
- A. Drieu et al., "Parenchymal border macrophages regulate the flow dynamics of the cerebrospinal fluid," *Nature* 611(7936), 585–593 (2022).
- J. Anrather and C. Iadecola, "Inflammation and stroke: an overview," *Neurotherapeutics* 13(4), 661–670 (2016).
- T. H. Sanderson et al., "Molecular mechanisms of ischemia-reperfusion injury in brain: pivotal role of the mitochondrial membrane potential in reactive oxygen species generation," *Mol. Neurobiol.* 47(1), 9–23 (2013).
- J. A. Zivin, "Acute stroke therapy with tissue plasminogen activator (tPA) since it was approved by the U.S. Food and Drug Administration (FDA)," *Ann. Neurol.* 66(1), 6–10 (2009).
- L. J. Millar et al., "Neonatal hypoxia ischaemia: mechanisms, models, and therapeutic challenges," *Front. Cell. Neurosci.* 11, 78 (2017).
- H. A. Choi, N. Badjatia, and S. A. Mayer, "Hypothermia for acute brain injury–mechanisms and practical aspects," *Nat. Rev. Neurol.* 8(4), 214–222 (2012).
- E. Gerace et al., "NIR laser photobiomodulation induces neuroprotection in an in vitro model of cerebral hypoxia/ischemia," *Mol. Neurobiol.* 58(10), 5383–5395 (2021).
- S. J. An, T. J. Kim, and B. W. Yoon, "Epidemiology, risk factors, and clinical features of intracerebral hemorrhage: an update," *J. Stroke* 19(1), 3–10 (2017).
- 217. T. S. van Solinge et al., "Fibrinolytics and intraventricular hemorrhage: a systematic review and meta-analysis," *Neurocritt. Care* **32**(1), 262–271 (2020).
- 218. A. Shanshool and V. V. Tuchin, "Recent advances in the laser radiation transport through the head tissues of humans and animals: a review," *J. Biomed. Photonics Eng.* **6**(4), 40201 (2021).
- M. Citron, "Alzheimer's disease: strategies for disease modification," *Nat. Rev. Drug Discov.* 9(5), 387–398 (2010).
- 220. R. Zhang et al., "Dose–effect relationships for PBM in the treatment of Alzheimer's disease," J. Phys. D Appl. Phys. 54(35), 353001 (2021).
- 221. B. R. Bloem, M. S. Okun, and C. Klein, "Parkinson's disease," Lancet 397(10291), 2284-2303 (2021).
- 222. R. M. A. de Bie et al., "Initiation of pharmacological therapy in Parkinson's disease: when, why, and how," *Lancet Neurol.* 19(5), 452–461 (2020).
- 223. L. M. L. de Lau and M. M. B. Breteler, "Epidemiology of Parkinson's disease," *Lancet Neurol.* 5(6), 525–535 (2006).
- A. H. V. Schapira, "Mitochondria in the aetiology and pathogenesis of Parkinson's disease," *Lancet Neurol.* 7(1), 97–109 (2008).
- 225. I. S. Martins et al., "Measurement of tissue optical properties in a wide spectral range: a review [Invited]," *Biomed. Opt. Express* **14**(1), 249–298 (2023).

- 226. V. Ganeshan et al., "Pre-conditioning with remote photobiomodulation modulates the brain transcriptome and protects against MPTP insult in mice," *Neuroscience* **400**, 85–97 (2019).
- 227. R. Dobson and G. Giovannoni, "Multiple sclerosis: a review," Eur. J. Neurol. 26(1), 27-40 (2019).
- R. J. Franklin and S. A. Goldman, "Glia disease and repair-remyelination," *Cold Spring Harbor Perspect. Biol.* 7(7), a020594 (2015).
- B. Roozenbeek, A. I. R. Maas, and D. K. Menon, "Changing patterns in the epidemiology of traumatic brain injury," *Nat. Rev. Neurol.* 9(4), 231–236 (2013).
- M. O. Blaya, A. P. Raval, and H. M. Bramlett, "Traumatic brain injury in women across lifespan," *Neurobiol. Dis.* 164, 105613 (2022).
- 231. V. Di Pietro et al., "Antioxidant therapies in traumatic brain injury," Antioxidants 9(3), 260 (2020).
- 232. A. C. McKee et al., "The neuropathology of chronic traumatic encephalopathy," *Brain Pathol.* 25(3), 350–364 (2015).
- P. W. Breen and V. Krishnan, "Recent preclinical insights into the treatment of chronic traumatic encephalopathy," *Front. Neurosci.* 14, 616 (2020).
- 234. J. LeMoult and I. H. Gotlib, "Depression: a cognitive perspective," Clin. Psychol. Rev. 69, 51-66 (2019).
- 235. G. S. Alexopoulos, "Mechanisms and treatment of late-life depression," *Transl. Psychiatry* 9(1), 188 (2019).
- M. A. Caldieraro and P. Cassano, "Transcranial and systemic photobiomodulation for major depressive disorder: a systematic review of efficacy, tolerability and biological mechanisms," *J. Affect. Disord.* 243, 262–273 (2019).
- P. Czarny et al., "The interplay between inflammation, oxidative stress, DNA damage, DNA repair and mitochondrial dysfunction in depression," *Progr. Neuropsychopharmacol. Biol. Psychiatry* 80, 309–321 (2018).
- 238. H. Zhao et al., "Comparative study on photobiomodulation between 630 nm and 810 nm LED in diabetic wound healing both *in vitro* and *in vivo*," *J. Innov. Opt. Health Sci.* 15(2), 2250010 (2022).
- Y. Tanaka et al., "Infrared radiation has potential antidepressant and anxiolytic effects in animal model of depression and anxiety," *Brain Stimul.* 4(2), 71–76 (2011).
- 240. I. Liguori et al., "Oxidative stress, aging, and diseases," Clin. Interv. Aging 13, 757-772 (2018).
- E. Kanasi, S. Ayilavarapu, and J. Jones, "The aging population: demographics and the biology of aging," *Periodontology* 2000 72(1), 13–18 (2016).
- B. A. Payne and P. F. Chinnery, "Mitochondrial dysfunction in aging: much progress but many unresolved questions," *Biochim. Biophys. Acta Bioenerg.* 1847(11), 1347–1353 (2015).
- 243. R. S. Sohal and M. J. Forster, "Coenzyme Q, oxidative stress and aging," *Mitochondrion* 7(Suppl.), S103–S111 (2007).
- 244. C. Grady, "The cognitive neuroscience of ageing," Nat. Rev. Neurosci. 13(7), 491-505 (2012).

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